

Improving coverage and consistency of MS-based proteomics

Limsoon Wong

(Joint work with Wilson Wen Bin Goh)



Proteomics vs transcriptomics

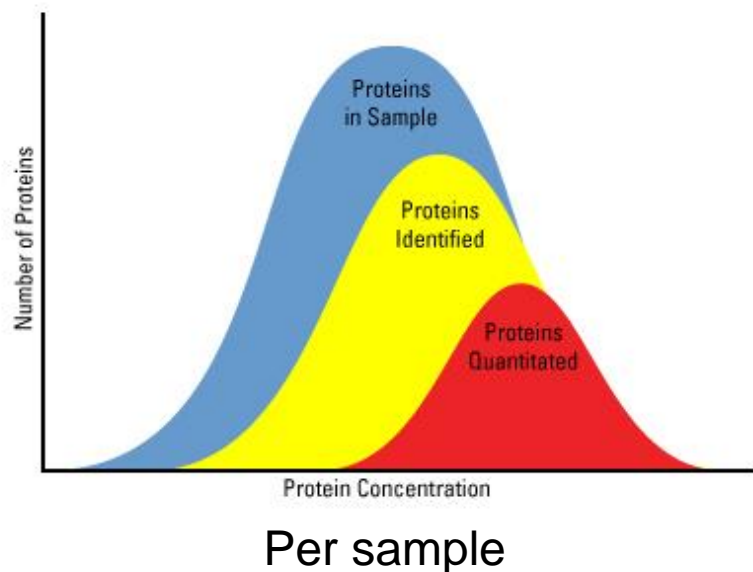


- **Proteomic profile**
 - Which protein is found in the sample
 - How abundant it is
- **Similar to gene expression profile. So typical gene expression profile analysis methods can be applied in theory...**
- **Key differences**
 - Profiling
 - **Complexity: 20k genes vs 500k proteins**
 - **Dynamic range: > 10 orders of magnitude in plasma. Proteins cannot be amplified**
 - Analysis
 - **Much fewer features**
 - **Difficult to reproduce**
 - **Much fewer samples**
 - **Unstable quantitation**

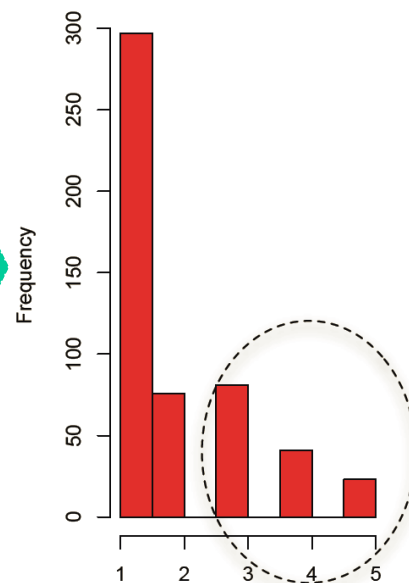
Issues in proteomics: Coverage and consistency

Technical incompleteness

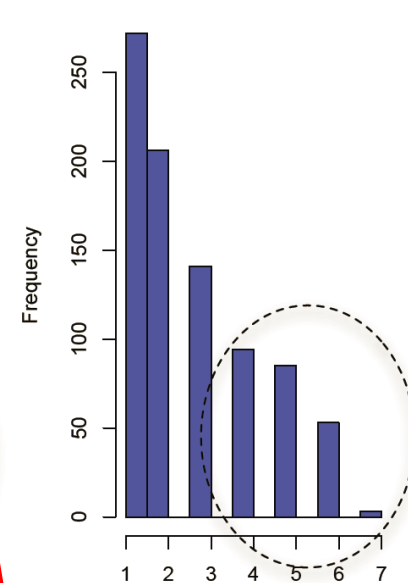
How it affects real data



Distribution of counts in mod



Distribution of counts in poor



Only 25 out of 800+ proteins are common to all 5 mod-stage HCC patients!

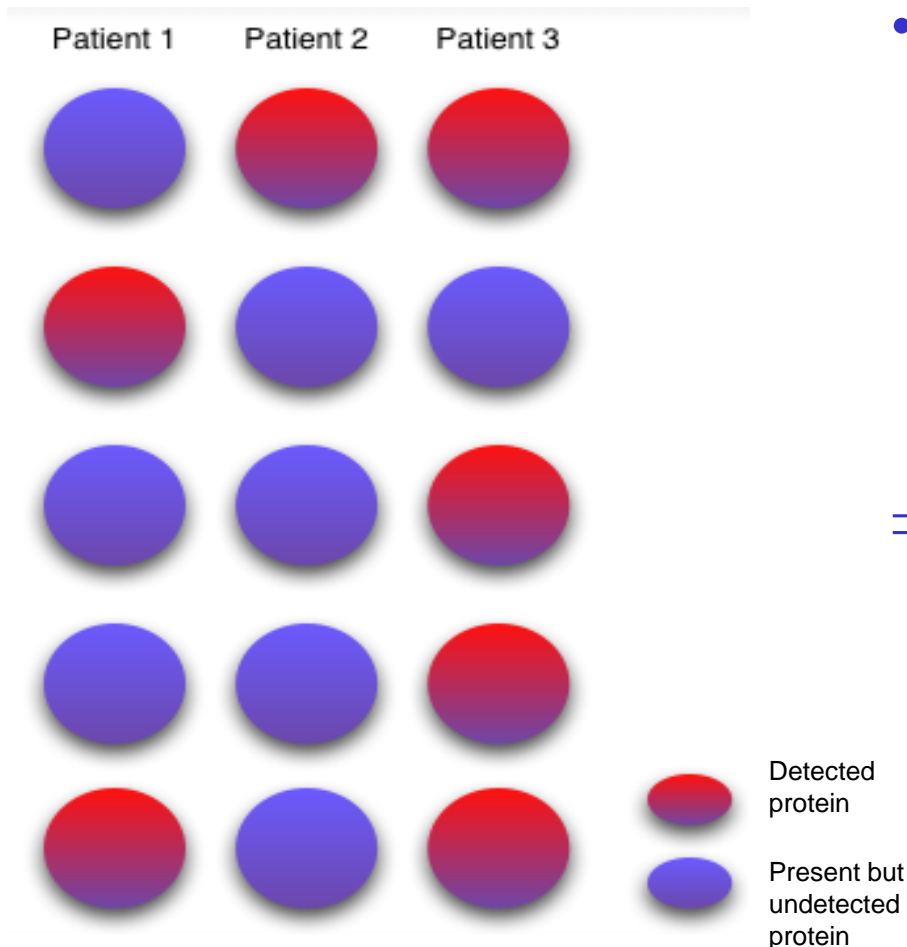
Using protein complexes to enhance proteomics: Basic ideas



A postulate and some math

- **Postulate: The chance of a protein complex being present in a sample is proportional to the fraction of its constituent proteins being correctly reported in the sample**
- **Suppose proteomics screen has 75% reliability; a complex comprises proteins A, B, C, D, E; and screen reports A, B, C, D only**
 - ⇒ **Complex has 60% ($= 0.75 * 4 / 5$) chance to be present**
 - ⇒ **The unreported protein E also has $\geq 60\%$ chance to be present, as presence of the complex implies presence of all its constituents**
 - ⇒ **improving coverage**
 - ⇒ **Each of the reported proteins (A, B, C, and D) individually has 90% ($= 100\% * 0.6 + 75\% * 0.4$) chance of being true positive, whereas a reported protein that is isolated has a lower 75% chance of being true positive**
 - ⇒ **removing noise**

An intuition



- **Suppose the failure to form a protein complex causes a disease**
 - If any component protein is missing, the complex can't form
- ⇒ **Diff patients suffering from the disease can have a diff protein component missing**
 - Construct a profile based on complexes?

Reference complexes

- In this talk, human complexes (of size at least 5) from CORUM are used as reference complexes
- It is possible to use subnets generated from pathway and PPI databases. However these such subnets vary significantly depending on network databases and subnet-generation algo used

So I do not
consider these...

Improving coverage in proteomic profiles



Lots of missing values in real proteomics datasets



nm.3807-S4.xls [Read-Only] [Compatibility Mode] - Microsoft Excel

	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T	U	V	W	X	Y	Z	AA	AB	AC			
	protein	GeneSy mbol	kidneyTisue1	kidneyTis ue2	kidneyTis ue3	kidneyTis ue4	kidneyTis ue5	kidneyTis ue6	kidneyTis ue7	kidneyTis ue8	kidneyTis ue9	kidneyTis ue10	kidneyTis ue11	kidneyTis ue12	kidneyTis ue13	kidneyTis ue14	kidneyTis ue15	kidneyTis ue16	kidneyTis ue17	kidneyTis ue18	kidneyTis ue19	kidneyTis ue20	kidneyTis ue21	kidneyTis ue22	kidneyTis ue23	kidneyTis ue24	kidneyTis ue25	kidneyTis ue26	kidneyTis ue27			
1	P09110	ACAA1	288001.7778	46353.28	237958.5	30102.47	37098.09	67454.84	92200.62	231528.4	12617.18	263299.1	NA	222387.2	NA	177211	27857.94	84689.84	43497.89	280540.3	77962.17	235242.5	23827.06	302761.4	41190.07	2064.747	97756.44	122386.3				
2	P05166	PCCB	246687.75	70504.27	253890.9	NA	314250.1	33680.65	108554.7	321442.7	260389.5	183399.7	258247.1	139288.5	284934.5	115138	245595.9	30488.41	221565	280540.3	340054.8	65477.99	250479.3	NA	327799	41974.24	1251043	321442.7	175808.5			
3	Q96RP9	GFM1	37872.59722	NA	40359.89	NA	73975.35	NA	64601.65	56815.28	34506.99	35176.2	98642.34	23060.3	91995.3	NA	37735.48	33491.8	48208.46	47858.24	39584.44	NA	67976.03	23631.74	46763.48	NA	2064.747	53619.99	67555.47			
4	Q15417	CNN3	28364.89722	NA	NA	NA	NA	44156.47	52272.02	27128.03	10577.49	32524.27	14171.12	33388.93	27593.38	49821.32	23144.21	24964.95	32403	NA	24907.94	46053.92	NA	NA	25129.86	42948.4	2064.747	26438.35	23207.51			
5	Q96FQ6	S100A16	NA	35176.2	NA	66058.39	NA	30674.6	1804.538	21706.65	NA	NA	11359.64	NA	18677.58	41493.97	12617.18	22496.77	NA	NA	NA	36422.79	NA	75858.83	20589.93	31161.06	2064.747	20398.13	NA			
6	P62820	RAB1A	NA	NA	NA	NA	NA	NA	54417.16	3130.811	NA	68503.39	NA	NA	NA	NA	NA	NA	NA	NA	32596.28	NA	NA	54839	NA	48748.28	2064.747	NA	NA			
7	P27169	PON1	NA	47101.83	58436.31	18128.35	NA	33573.36	112930.6	NA	NA	NA	59432.1	NA	39084.55	36282.92	16953.34	NA	NA	NA	45107.13	NA	19506.67	NA	38130.55	190838.9	NA	NA				
8	Q9UL46	PSME2	33680.65278	99968.93	59047.33	145114.2	33256.26	141575.7	77962.17	75727.38	64365.04	121022.2	40286.83	40567.01	104458.4	42876.78	83666.14	55954.92	62742.03	33768.27	111940.8	59915.42	151558.9	38443.16	113145.5	79024.33	73747.38	40140.37				
9	P08237	PFKM	39644.09722	NA	54240.61	NA	136064	NA	1804.538	62845.97	141296.3	100616.3	137596.7	NA	140860.9	NA	96590.73	NA	92823.65	51085.24	155550.8	NA	47697.29	NA	136064	NA	2064.747	58618.05	143381.1			
10	P04040	CAT	292456.0528	149632.6	239229.2	24964.95	528247.1	220764.4	540115.8	133921.9	289434.5	367784.7	293727.3	179981.9	259314.6	14294.3	204722.1	77070.33	109006.7	136875.9	290924.4	163095.2	237958.5	31389.75	271920.4	22790.3	49942.8	150524.5	294964.3			
11	Q8WYA6	CNNB1	NA	NA	NA	NA	NA	NA	1804.538	NA	NA	NA	NA	NA	NA	NA	27646.1	37621.73	26686.24	NA	NA	NA	NA	NA	NA	NA	2064.747	NA	NA			
12	Q9H0W9	C11orf54	454591.5833	77225.75	393512.7	55431.72	365975.5	180535.1	188742.5	77348.17	352898.9	119242.7	417999.9	263299.1	474797	229655.9	427428	143697	124568	146454.4	441856.5	74156.41	370040.5	44605.86	363784.6	187566.8	129074.8	104101.6	375463.4			
13	P13198	STIP1	76018.00556	83236.9	83516.5	137596.7	75613.89	110367.2	98642.34	195146	77709.53	282315.9	65948.94	122386.3	81635.42	129969.2	67749.81	124568	108554.7	135737.2	69039.96	92656.4	85600.47	147792.9	65262.99	109273.7	91127.04	218888	122047.2			
14	O59401	SUN1	57623.33889	NA	NA	NA	72273.86	NA	1804.538	NA	NA	NA	80603.49	NA	NA	NA	NA	NA	NA	NA	60013.66	NA	NA	71252.19	NA	2064.747	NA	NA				
15	Q97174	HSD17B10	175372.7444	114480.8	181096.8	75400.28	222387.2	91466.47	218888	269679.7	179177.4	165285.9	202618.2	117389.5	191537	41135.21	196208.5	151044.7	210269.6	294964.3	183893	82644.38	179981.9	102286.8	233372.9	91325.89	196968.8	293727.3	174540.8			
16	Q15833	STXB2	14224.84722	24264.99	14303.05	19690.86	16316.33	NA	1804.538	NA	14303.05	17309.98	11459.84	14224.85	12617.18	NA	14224.85	9837.458	21131.38	5634.228	13283.71	28846.59	20057.06	12924.71	17380.49	NA	2064.747	11880.63	13166.66			
17	P08195	SLC3A2	50797.625	42825.82	63302.14	26628.24	85345.18	NA	1804.538	NA	77850.57	NA	100616.3	NA	76579.02	NA	44010.16	17146.31	17146.31	NA	80199.58	41362.6	72273.86	32198.97	75858.83	NA	2064.747	NA	76292.57			
18	P26038	MSN	333342.6833	438752.3	421056.2	381249.5	241992.3	404349.8	163434.5	172028.6	446678.9	167923.7	367784.7	310472.5	404349.8	393512.7	292456.1	427428	390317.5	244865.7	273261.7	446678.9	404349.8	306071.8	222387.2	423963.5	191537	182241.6	441856.5			
19	P09104	ENO2	NA	144058.2	NA	184650.5	NA	137596.7	12616.3	21831.56	NA	NA	NA	NA	NA	404349.8	NA	48438.29	57080.76	NA	151558.9	NA	181096.8	NA	123793.9	2064.747	NA	NA				
20	P07148	FABP1	1219163.714	34579.48	861796.3	NA	940142	NA	1804.538	NA	1130692	NA	1057986	NA	789446.1	NA	221565	NA	NA	NA	1162786	32336.43	805128.4	NA	970053.3	NA	2064.747	NA	1300718			
21	Q96Q11	TRNT1	NA	NA	NA	NA	NA	NA	1804.538	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	37098.09	35565.03	NA	NA	NA	NA	NA	2064.747	NA	NA			
22	O15083	ERC2	NA	NA	NA	85740.42	NA	NA	1804.538	NA	83390.33	NA	NA	NA	NA	NA	NA	NA	142306.8	NA	NA	NA	72396.48	NA	NA	2064.747	NA	70213.43				
23	Q15911	ZFH3	NA	178745.3	393512.7	205865.1	682653.9	1804.538	NA	243050.1	NA	189860.5	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	2064.747	NA	252846.2				
24	Q9BUR5	APOO	35479.70278	NA	27260.11	15459.06	40140.37	NA	1804.538	46154.89	30730.15	54737.36	47185.33	13642.38	28517.17	NA	40140.37	NA	NA	10649.17	34436.2	NA	36956.08	16653.18	47858.24	2064.747	33003.64	20057.06				
25	Q9UI83	HACL1	417999.9306	NA	435248.4	NA	336790.8	227161.7	1804.538	174111.8	276628.6	NA	274264.6	NA	317227.1	271920.4	336790.8	NA	NA	372485.6	446678.9	NA	390317.5	NA	307205	211073.8	2064.747	169817.6	33342.7			
26	Q8WU44	PDCD6IP	50008.50556	34991.44	70504.27	50108.55	59047.33	41611.18	84319.78	97140.59	56715.96	134561.7	52110.31	61553.77	67555.47	65262.99	68597.03	59827.38	73200.35	75044.44	64108.37	40359.89	79093.29	49636.31	49821.32	37258.59	76579.02	76865.11	37386.23			
27	P53597	SUGL1	387432.1583	99433.589	228946.3	94932.09	310472.5	150524.5	187002.3	299487.5	275420.7	308775.7	299487.5	101732.7	245595.9	108554.7	270810.9	89524.72	192915.6	276628.6	357417.6	96739.9	205171.6	95793.82	288001.8	162300.5	193664.8	299487.5	245595.9			
28	O00186	STXB3	NA	28468.21	NA	NA	NA	19019.68	1804.538	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	15575.29	29005.53	NA	NA	NA	2064.747	NA	NA				
29	Q8N335	GPDI1	52415.71111	NA	59328.51	NA	54240.61	21949.83	109838.9	91466.47	54227.61	109273.7	50443.03	NA	52700.48	23221.01	45502.32	NA	57623.34	41362.6	54737.36	NA	62380.69	NA	93224.91	54839	23827.06	152627.3	71658.52	49636.31		
30	P08621	SNRNP70	48594.65	51791.05	47269.07	86082.28	44306.32	53026.19	1804.538	NA	59432.1	54839	49636.31	60605.33	52477.21	NA	72977.35	74546.25	82242.07	33003.64	60605.33	49636.31	93224.91	NA	56917.54	2064.747	NA	50797.63				
31	Q969V6	MKL1	NA	91325.89	55594.92	NA	74269.09	80102.57	1804.538	NA	71906.43	NA	NA	152627.3	72497.5	72497.5	89662.88	51690.71	68707.95	41576.85	72021.55	92973.8	NA	NA	NA	88904.66	2064.747	NA	NA			
32	P08311	CTSG	NA	NA	46154.89	NA	NA	67879.78	1804.538	NA	53026.19	NA	NA	68927.99	NA	NA	NA	NA	218057.1	78414.15	NA	NA	46895.88	NA	NA	56514.53	66379.24	NA	NA			
33	Q9UKU7	ACAD8	46053.91944	31797.32	50179.16	NA	64601.65	NA	75160.02	49228.15	440																					

Missing values
are not due
mostly to low-
abundance
proteins

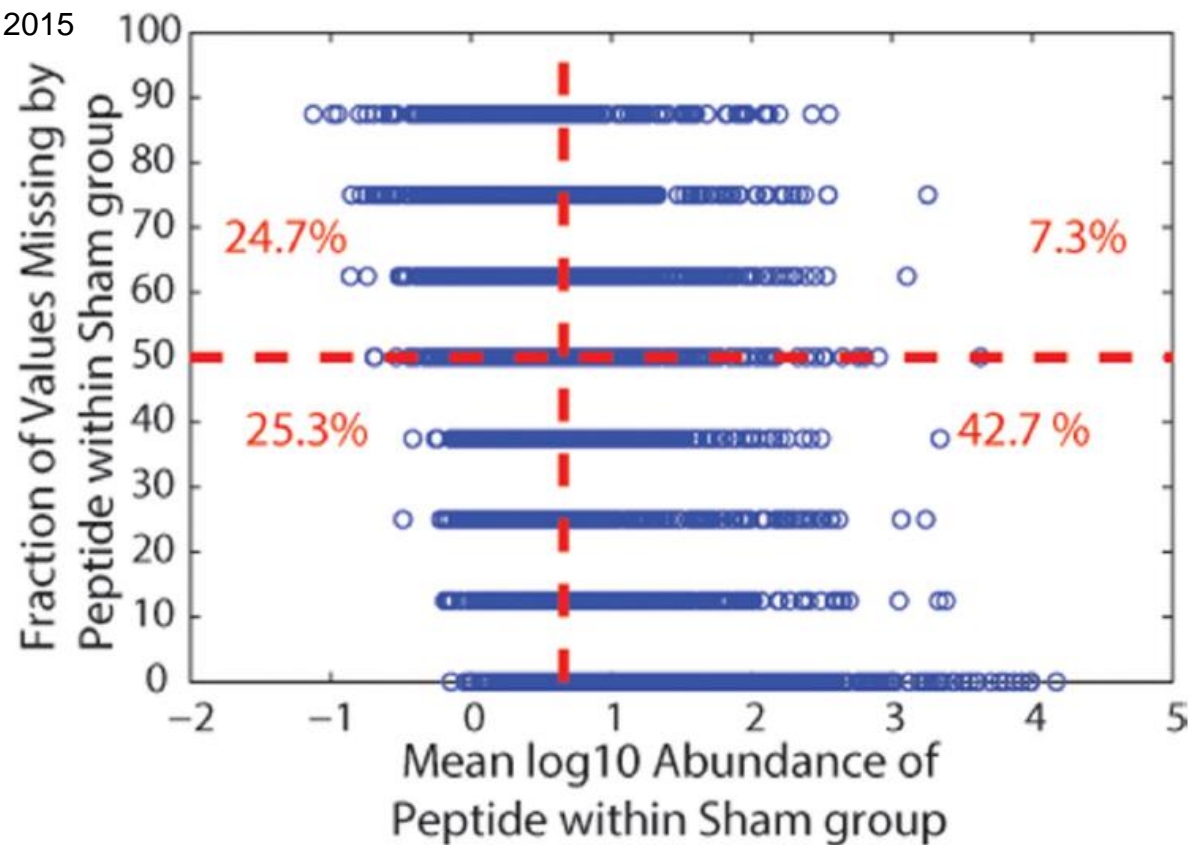


Figure 1.

Average \log_{10} intensity as measured by peptide peak area in the control group versus fraction of missing values and peptide counts associated with bins corresponding to the fraction of missing data comparing phenotypes and exposures for datasets from (A) human plasma and (B) mouse lung. The control group for the human plasma is the normal glucose tolerant (NGT) samples, and the sham group for the mouse lung is the regular weight mice with no lipopolysaccharide (LPS) exposure. The vertical red line represents median average intensity, and the horizontal red line represents the point that 50% of the values are missing.

Current
 imputation
 methods
 don't work
 very well

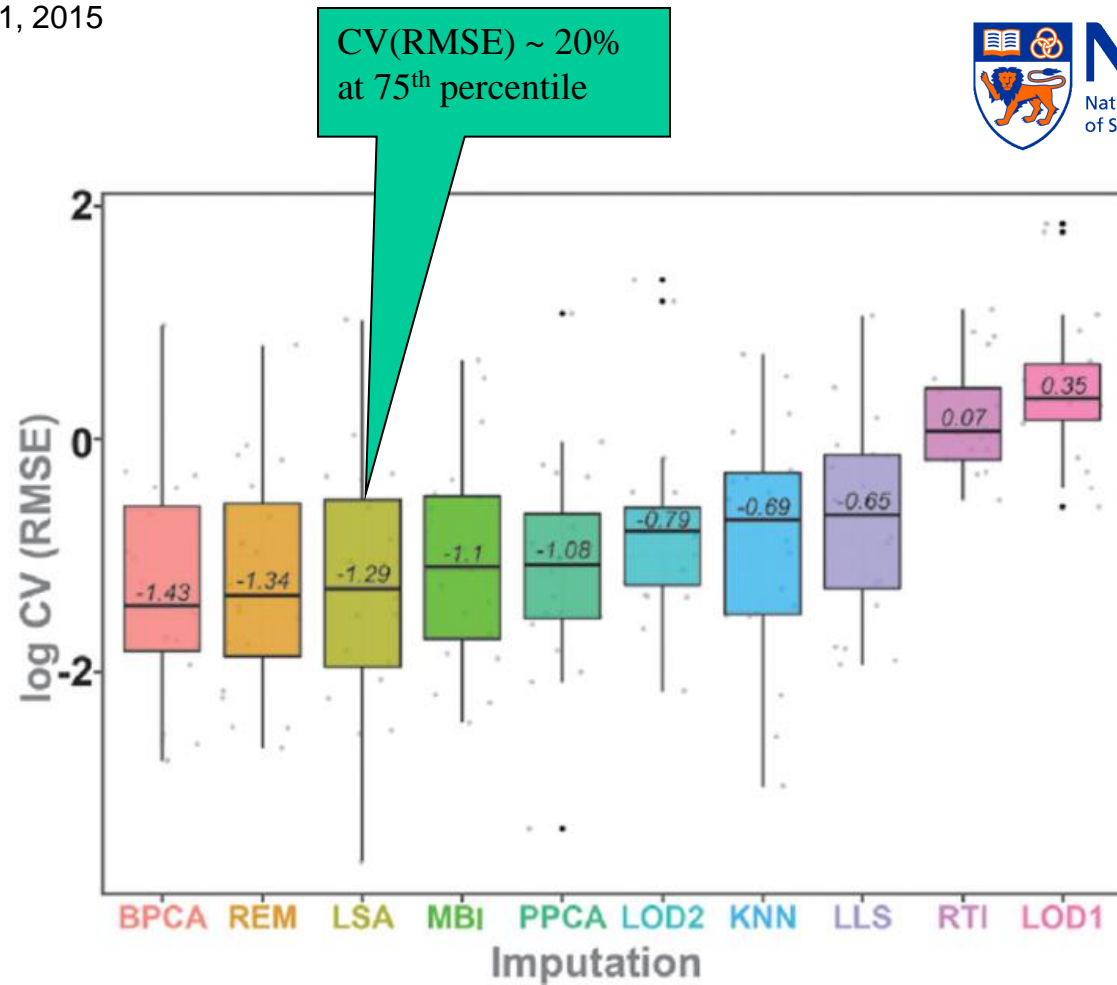
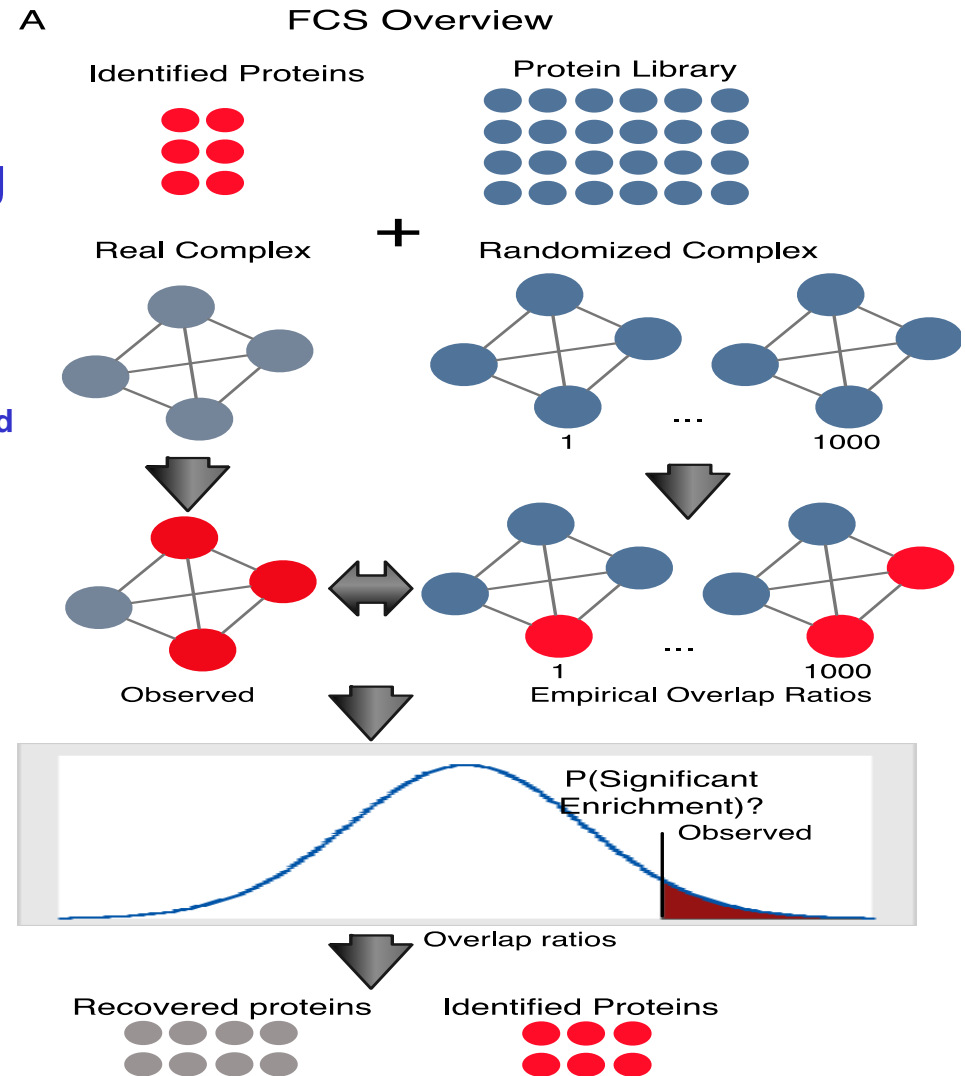


Figure 2.

Boxplot of the average $\log_{10} \text{CV(RMSE)}$ for the imputed dilution series datasets (Table 1) at the (A) peptide and (B) protein levels. The lower line represents the 25th percentile, the upper line of the box represents the 75th percentile, and the inner line corresponds to the median $\log_{10} \text{CV(RMSE)}$.

FCS

- **Rescue undetected proteins from high-scoring protein complexes**
- Goh et al. Comparative network-based recovery analysis and proteomic profiling of neurological changes in valproic acid-treated mice. *JPR*, 12(5):2116-2127, 2013



Other methods for rescuing missing proteins



- **CEA**

- Generate cliques from PPIN
- Rescue missing proteins from cliques containing lots of high-confidence proteins
- Li et al. Network-assisted protein identification and data interpretation in shotgun proteomics. *Mol. Syst. Biol.*, 5:303, 2009

- **MaxLink**

- Map high-confidence proteins (“seeds”) to PPIN
- Rescue proteins that interact many seeds but few non-seeds
- Goh et al. *Int J Bioinformatics Research and Applications*, 8(3/4):155-170, 2012

- **PEP**

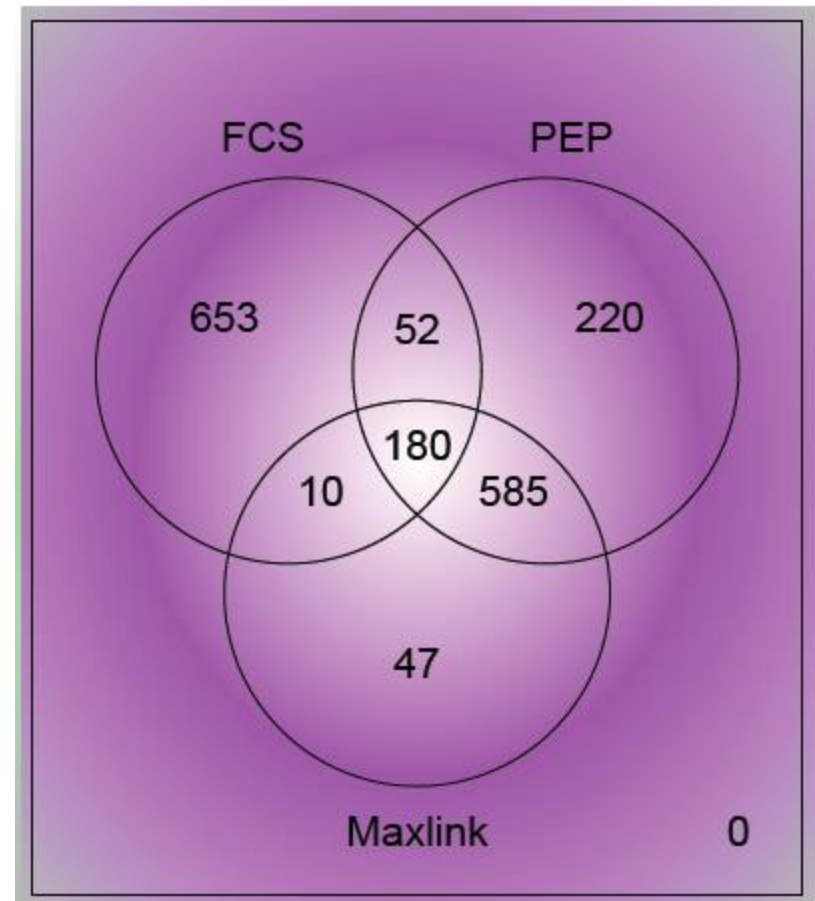
- Map high-confidence proteins to PPIN
- Extract neighbourhood & predict protein complexes using CFinder
- Rescue undetected proteins from high-ranking predicted complexes
- Goh et al. A Network-based pipeline for analyzing MS data---An application towards liver cancer. *J. Proteome Research*, 10(5):2261-2272, 2011

iTRAQ experiment

- **Valporic acid (VPA)-treated mice vs control**
 - VPA or vehicle injected every 12 hours into postnatal day-56 adult mice for 2 days
 - Role of VPA in epigenetic remodeling
- **MS was scanned against IPI rat db in round #1**
 - 291 proteins identified
- **MS was scanned against UniProtkb in round #2**
 - 498 additional proteins identified
- **All recovery methods ran on round #1 data and the recovered proteins checked against round #2**

Moderate level of
agreement of
reported proteins
between various
recovery methods

FCS (Real Complexes)



Performance comparison

Method	Novel Suggested Proteins	Recovered proteins	Recall	Precision
PEP	1037	158	0.317	0.152
Maxlink	822	226	0.454	0.275
FCS (predicted)	638	224	0.450	0.351
FCS (complexes)	895	477	0.958	0.533

- Looks like running FCS on real complexes is able to recover more proteins and more accurately

SWATH experiment

- **If there are technical replicates, they should have reported the same proteins. So we can run FCS on one replica, and see whether the predicted missing proteins show up in other replicas**
- **If there are multiple biological replicates (i.e. patients of the same phenotype), we can run FCS on one of them, and check on the others**
- **Proteomics data used: Renal cancer**
 - Guo et al. *Nature Medicine*, 21(4):407-413, 2015
 - 6 pairs of normal vs cancer ccRCC tissues
 - SWATH in duplicates

~20% of predicted missing proteins
 are supported by ≥ 1 reported
 peptide in the screen

A Strategy 1 (complex to proteins in the peptide list back to self)

Sample	N T1-> N T1	N T2 -> N T2	C T1-> C T1	C T2 -> C T2
1	0.203 0 985 200	0.220 0 937 206	0.186 0.001 823 153	0.191 0 911 174
2	0.204 0 936 191	0.222 0 889 197	0.194 0.004 904 175	0.215 0 918 197
3	0.197 0 972 191	0.212 0 950 201	0.241 0 849 205	0.225 0 840 189
4	0.223 0 943 210	0.232 0 948 220	0.215 0.001 925 199	0.211 0 930 196
5	0.225 0 912 205	0.201 0 964 194	0.209 0 877 183	0.185 0 904 167
6	0.249 0 883 220	0.215 0 977 210	0.233 0 886 206	0.241 0 927 223

~20% of predicted missing proteins
are supported by ≥ 1 reported
peptide in the replicate



B Strategy 2 (complex to proteins in the peptide list in the other replicate)

Sample	N T1-> N T2	N T2 -> N T1	C T1-> C T2	C T2 -> C T1
1	0.212 0 985 209	0.210 0 937 197	0.198 0 823 163	0.182 0 911 166
2	0.213 0 936 199	0.216 0 889 192	0.205 0 904 185	0.202 0.001 918 185
3	0.212 0 972 206	0.196 0 950 186	0.218 0 849 185	0.249 0 840 209
4	0.224 0 943 211	0.233 0 948 221	0.197 0.002 925 182	0.222 0 930 206
5	0.188 0.002 912 171	0.235 0 964 227	0.185 0 877 162	0.209 0 904 189
6	0.224 0 883 198	0.246 0 977 240	0.227 0 886 201	0.249 0 927 231

But ~25% of predicted missing proteins are supported by peptides in the screen or replicate



C Strategy 3 (complex to proteins in the peptide list union of self and other replicate)

Sample	N T1-> N T12	N T2 -> N T12	C T1-> C T12	C T2 -> C T12
1	0.248 0 985 244	0.258 0 937 242	0.238 0 823 196	0.229 0.001 911 209
2	0.248 0 936 232	0.260 0 889 231	0.225 0 904 203	0.234 0.001 918 215
3	0.243 0 972 236	0.241 0 950 229	0.274 0 849 233	0.281 0 840 236
4	0.268 0 943 253	0.280 0 948 265	0.251 0 925 232	0.263 0 930 245
5	0.254 0 912 232	0.267 0 964 257	0.241 0 877 211	0.238 0 904 215
6	0.280 0 883 247	0.275 0 977 269	0.269 0 886 238	0.283 0 927 262

~25% FCS-predicted missing proteins
 are supported by peptides in screen/replicate.
 Can we do better?

Recall this postulate:

The chance of a protein complex being present is proportional to the fraction of its protein members being correctly reported in the screen

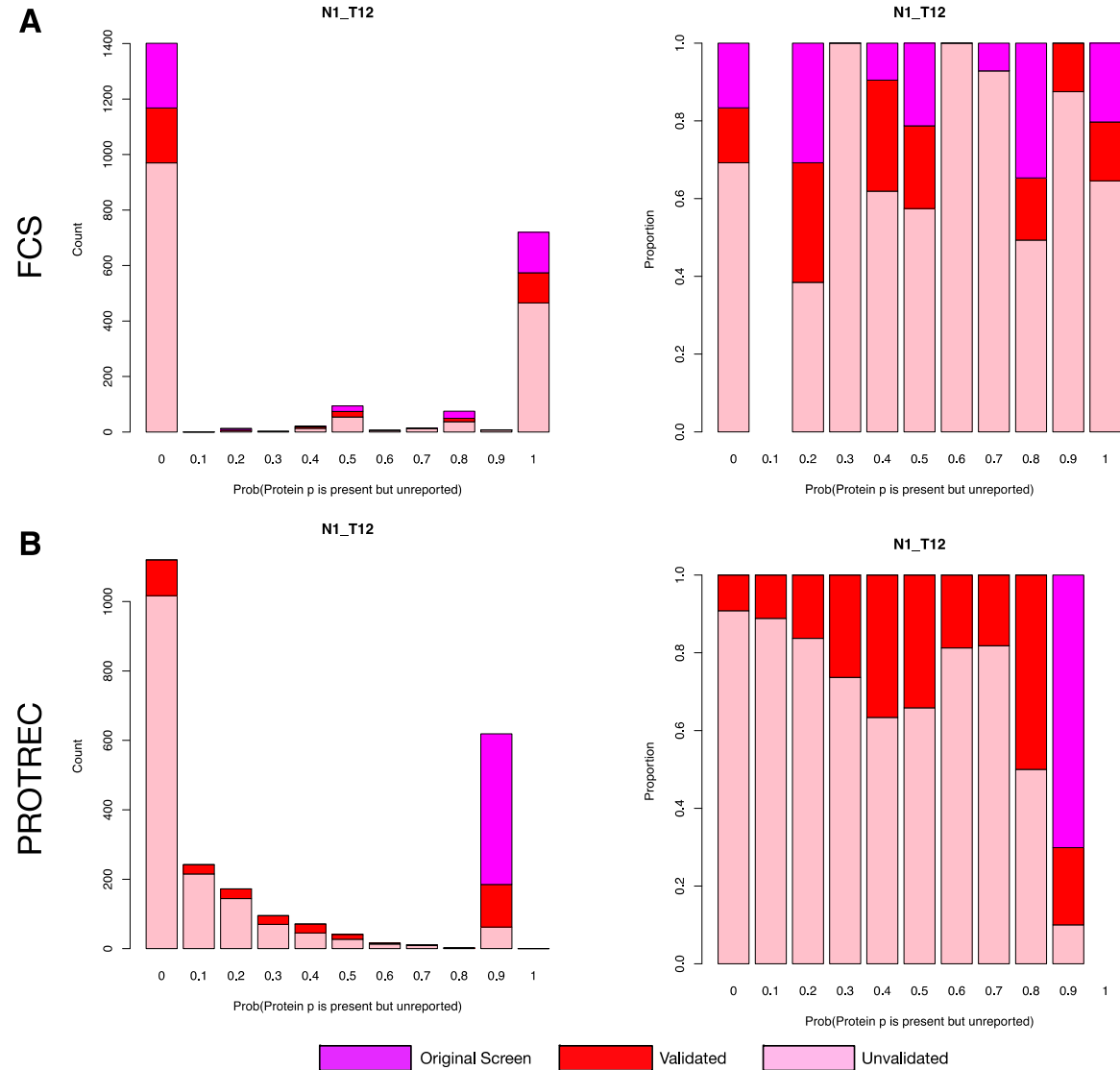
**Presence of complex implies
 presence of all member proteins**

PROTREC: Rank predicted missing proteins by

Prob(Protein p is present but unreported) =

$$\text{Max}_{\text{complex } C \text{ contains } p} \text{Prob}(p \text{ is present} \mid C \text{ is present}) * \text{Prob}(C \text{ is present}) + \text{Prob}(p \text{ is present} \mid C \text{ is absent}) * \text{Prob}(C \text{ is absent})$$

Ranking by
 PROTREC
 significantly
 improves
 precision of
 FCS
 predictions



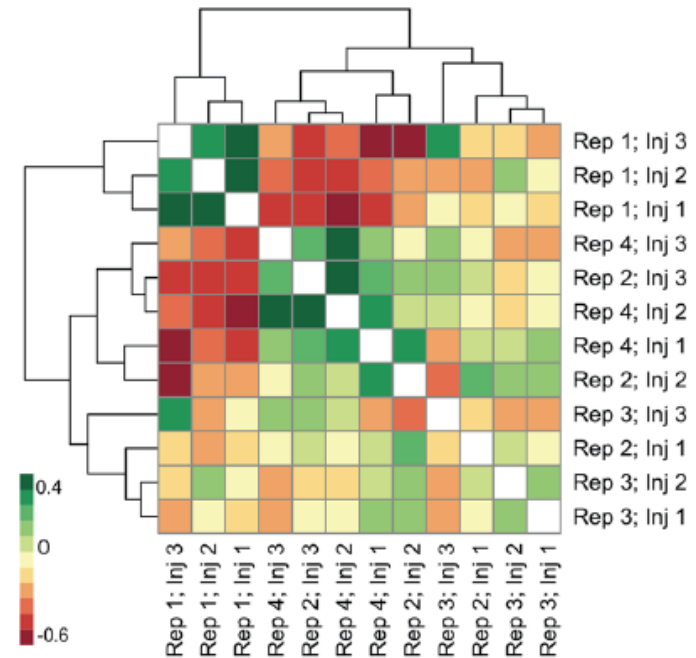
Improving consistency in proteomic profile analysis



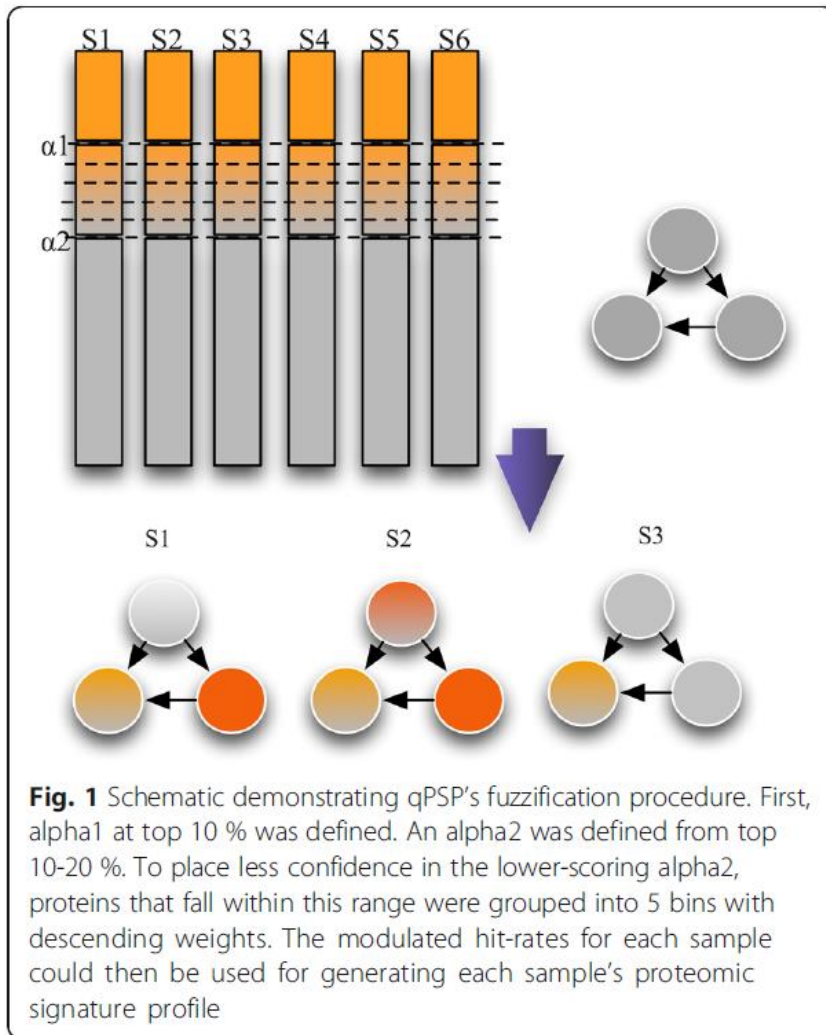
Proteomic profiles generally not consistent, even for technical replicates



- **A human kidney tissue**
 - Guo et al. *Nature Medicine*, 21(4):407-413, 2015
 - Digested in quadruplicates
 - Analyzed in triplicates
- **Clustering by proteins**
 - Correlation betw replicates is not good (~ 0.4)
 - Technical replicates of the same biological replicate are not tightly clustered



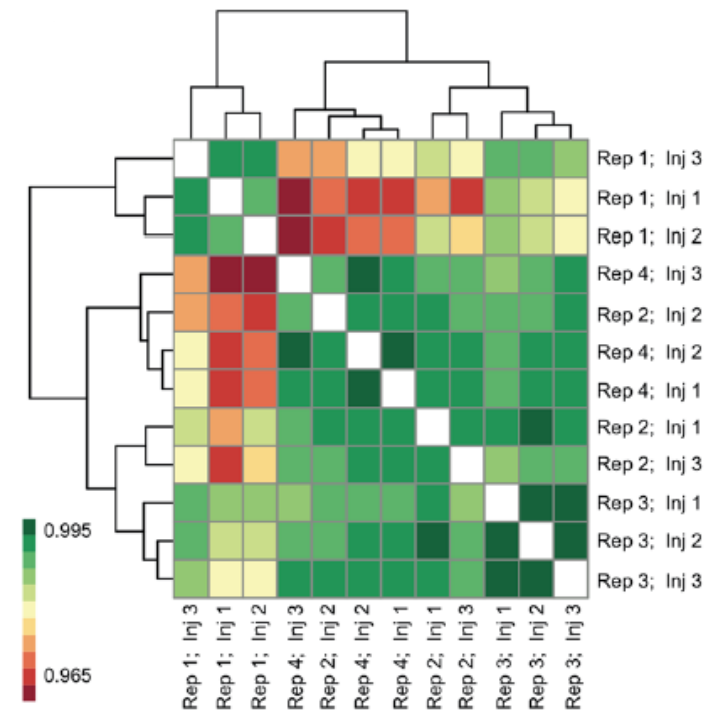
qPSP



- **Features are complexes**
- **Feature values are fuzzy weighted proportion of proteins in a complex**
 - $\text{score}(C, S_i) = \sum_{p \in C} \text{fs}(p, S_i) / |C|$
- **Complex C is significant if $\{\text{score}(C, S_i) \mid S_i \in A\}$ is very different by t-test from $\{\text{score}(C, S_i) \mid S_i \in B\}$**

Consistency of qPSP

- **Clustering of benchmarking control data based on protein complexes (i.e. qPSP)**
 - Correlation betw replicates is >0.95
 - **Cf. 0.4 based on proteins**
 - Technical replicates are better clustered



Further improving consistency, as well as
catching significant low-abundance
complexes



ESSNet, adapted for proteomics

- Let g_i be a protein in a given protein complex
- Let p_j be a patient
- Let q_k be a normal
- Let $\Delta_{i,j,k} = \text{Expr}(g_i, p_j) - \text{Expr}(g_i, q_k)$
- Test whether $\Delta_{i,j,k}$ is a distribution with mean 0

- Null hypothesis is “Complex C is irrelevant to the difference between patients and normals, and the proteins in C behave similarly in patients and normals”
- No need to restrict to most abundant proteins
- ⇒ Potential to reliably detect low-abundance but differential proteins

Lim et al. **A quantum leap in the reproducibility, precision, and sensitivity of gene expression profile analysis even when sample size is extremely small.** *JBCB*, 13(4):1550018, 2015

Five methods to compare with

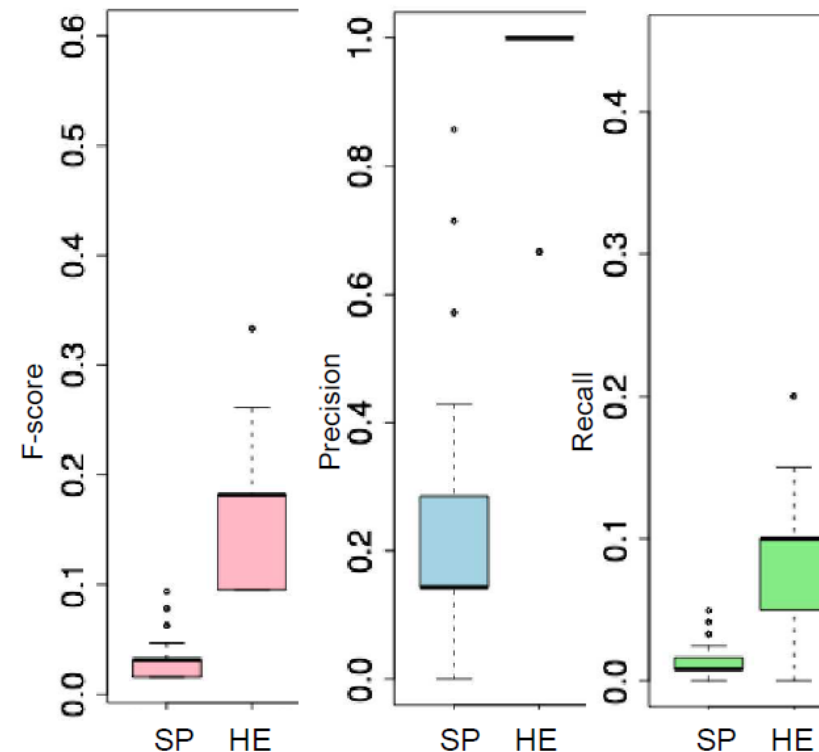
- **Network-based methods**
 - Hypergeometric enrichment (HE)
 - Direct group analysis (DG), similar to GSEA
 - qPSP, Goh et al., *Biology Direct*, 10:71, 2015
 - PFSNET, Goh & Wong, *JBCB*, 14(5):16500293, 2016
- **Standard t-test on individual proteins (SP)**

Simulated data

- **Simulated datasets from Langley and Mayr**
 - D.1.2 is from study of proteomic changes resulting from addition of exogenous matrix metallopeptidase (3 control, 3 test)
 - D2.2 is from a study of hibernating arctic squirrels (4 control, 4 test)
- **Both D1.2 and D2.2 have 100 simulated datasets, each with 20% significant features**
 - Effect sizes of these differential features are sampled from one out of five possibilities (20%, 50%, 80%, 100% and 200%), increased in one class and not in the other
- **Significant artificial complexes are constructed with various level of purity (i.e. proportion of significant proteins in the complex)**
 - Equal # of non-significant complexes are constructed as well

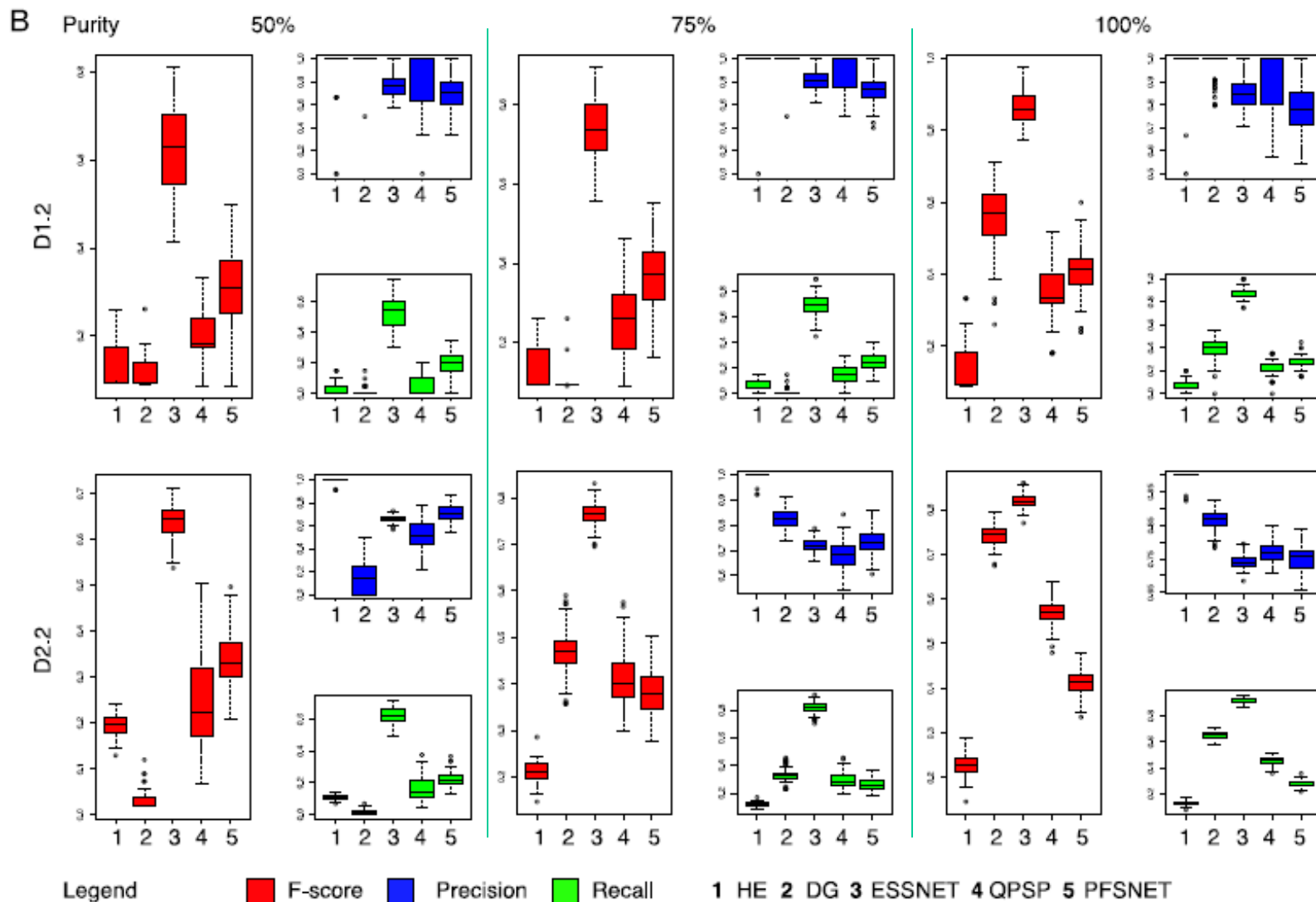
SP shows poor performance on simulated data.

Can network-based methods do better?



Supplementary Figure 1 Single protein (SP) precision-recall performance on D1.2. The f-score (pink), precision (blue) and recall (green) shows that SP performs abysmally on simulated data. HE is shown next to SP as a reference.

ESSNET shows excellent recall/precision on simulated data

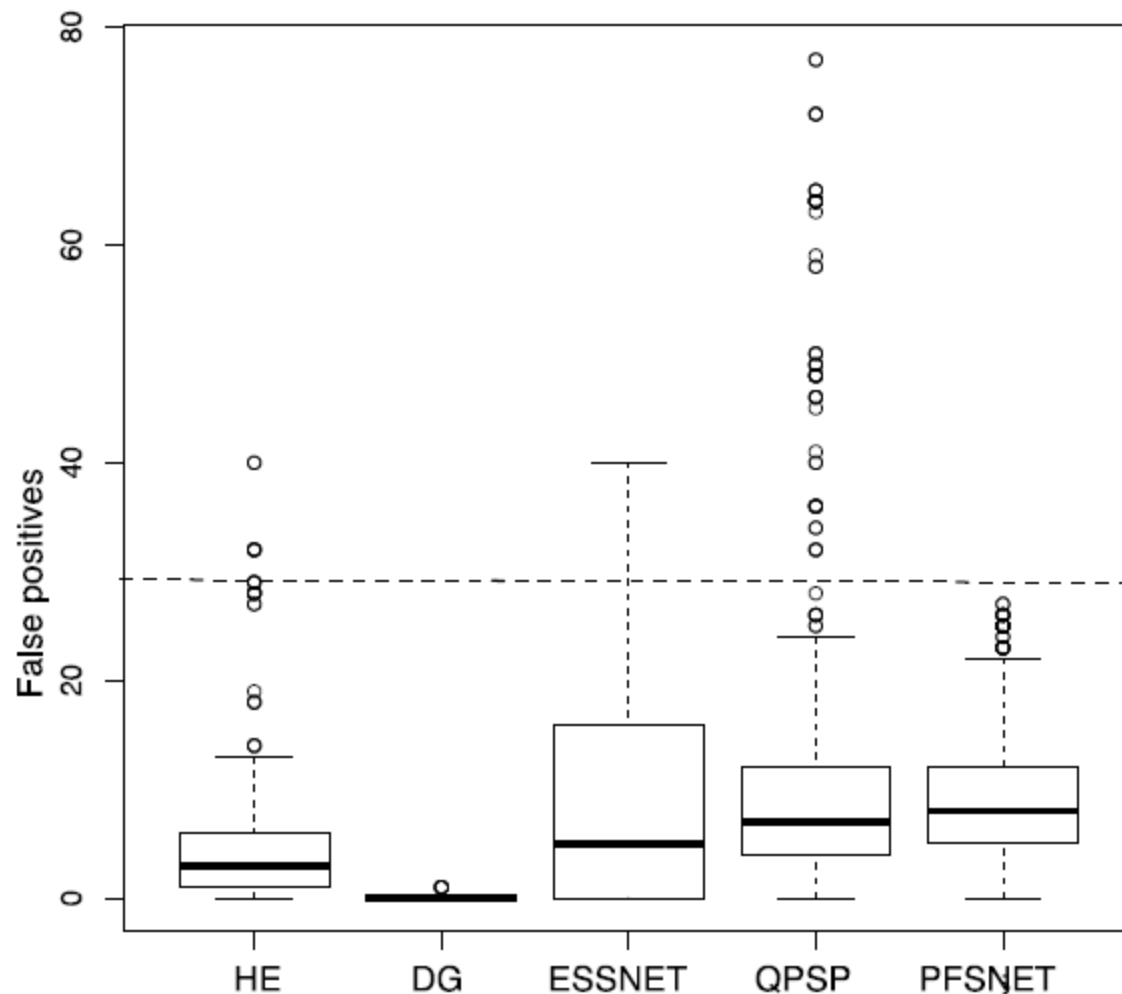




Renal cancer control data (RCC)

- **12 runs originating from a human kidney tissue digested in quadruplicates and analyzed in triplicates**
- **Excellent for evaluating false-positive rates of feature-selection methods**
 - Randomly split the 12 runs into two groups. Report of any significant features between the groups must be false positives

All
 methods
 control
 false
 positives
 well



Dash line corresponds to expected # of false positives at alpha 0.05 (~30 complexes)

Renal cancer data (RC)

- **12 samples are run twice so that we have technical replicates over 6 normal and 6 cancer tissues**
- **Excellent opportunity for testing reproducibility of feature-selection methods**
 - A good method should report similar feature sets between replicates
- **Can also test feature-selection stability**
 - Apply feature-selection method on subsamples and see whether the same features get selected

ESSNET & PFSNET show excellent reproducibility

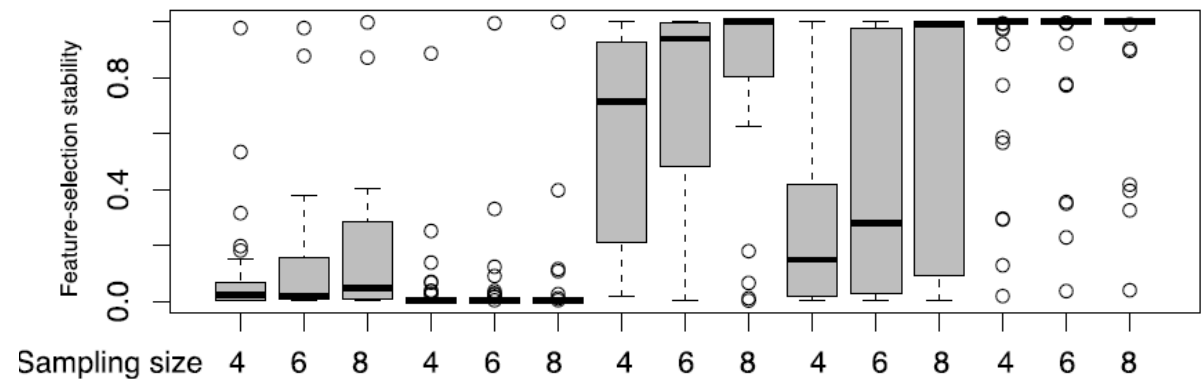
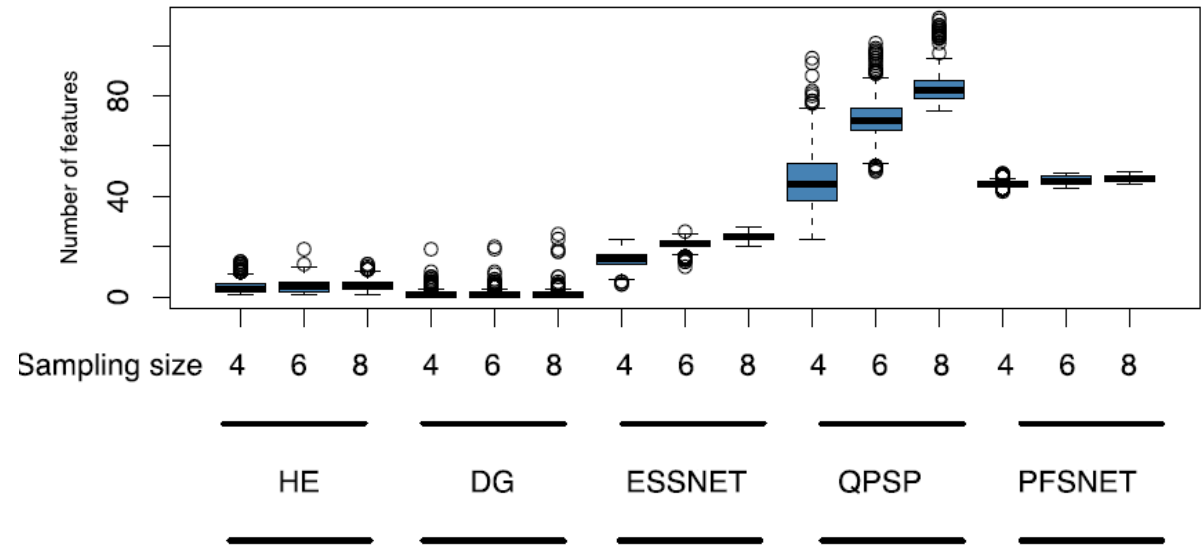


Number of terms	HE	DG	ESSNET	QPSP	PFSNET
Replicate 1	4	1	35	86	45
Replicate 2	6	2	29	75	46
Overlaps	0.25	0.5	0.83	0.66	0.94

HE	DG	ESSNET	QPSP	PFSNET	
1	0.5	0.71	0.86	0.71	HE
	1	1	1	1	DG
		1	0.93	0.98	ESSNET
			1	0.90	QPSP
				1	PFSNET

This table is computed on by applying the methods on the full RC dataset

ESSNET &
 PFSNET
 show
 excellent
 stability



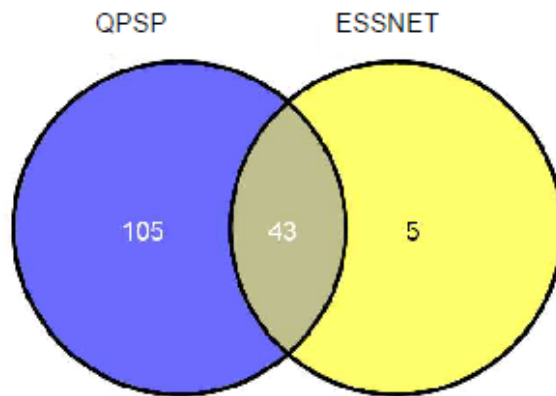
ESSNET &
 PFSNET
 show
 excellent
 stability

	4	6	8	Mean
HE	0.022	0.016	0.047	0.030
DG	0.001	0.001	0.002	0.001
ESSNET	0.714	0.941	1.000	0.885
QPSP	0.149	0.282	0.991	0.470
PFSNET	1.000	1.000	1.000	1.000

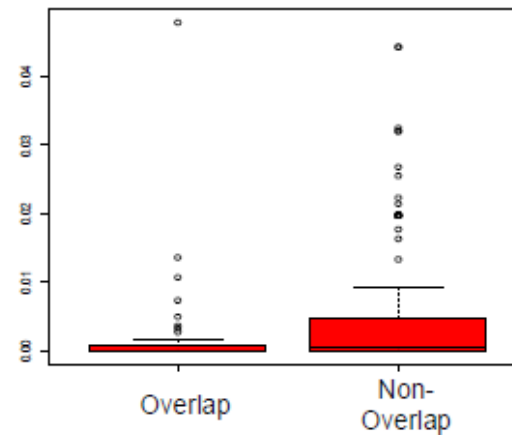
ESSNET can assay low-abundance complexes that qPSP cannot



A



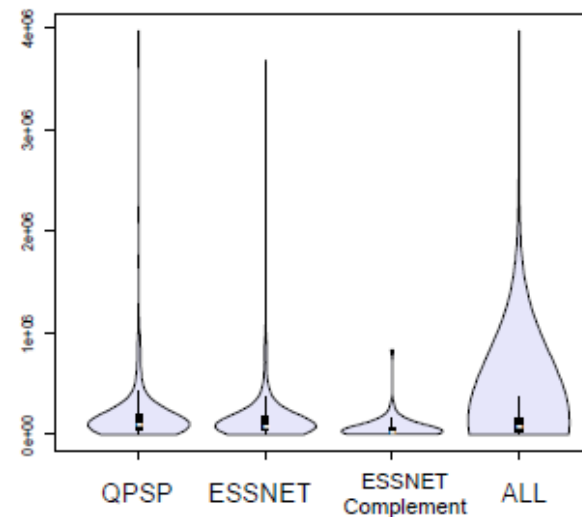
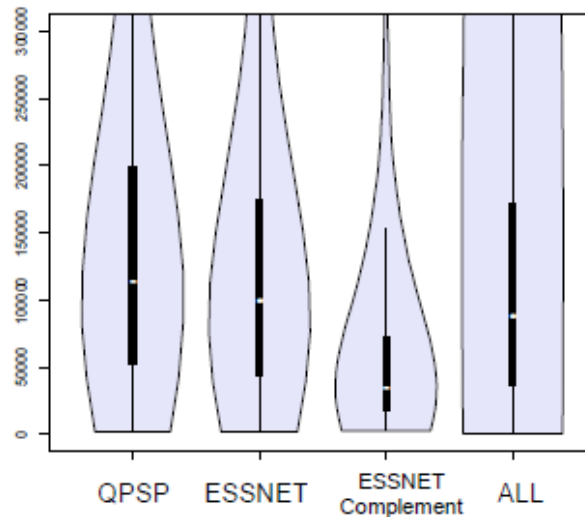
B



A: QPSP-ESSNET significant-complex overlaps

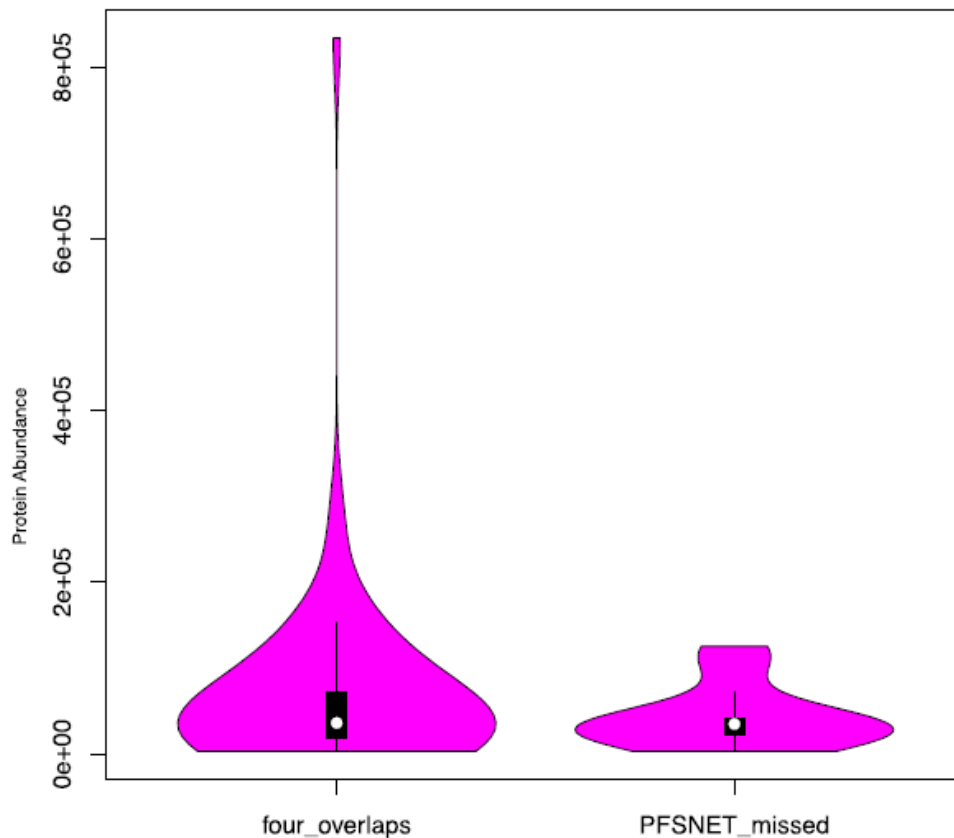
B: P-value distribution for overlapping and non-overlapping QPSP complexes.

C



C: Sampling abundance distribution. The left panel is a zoom-in of the right. The y-axis is the protein abundance while the four categories are the distribution of abundances of complexes found in QPSP, ESSNET, ESSNET unique (complement), and all proteins in RC.

ESSNET can assay low-abundance complexes that PFSNET cannot



Of the 5 ESSNET-unique complexes, PFSNET can detect 4; the missed complex consists entirely of low-abundance proteins.

If p-value threshold is adjusted by Benjamini-Hochberg 5% FDR, PFSNET can detect only 3 of the 5 ESSNET-unique complexes while ESSNET continues to detect them all.

Concluding Remarks



In conclusion...

Contextualization (into complexes) can deal with coverage and consistency issues in proteomics

References

- Goh & Wong. **Integrating networks and proteomics: Moving forward.** *Trends in Biotechnology*, in press
- [FCS] Goh et al. **Comparative network-based recovery analysis and proteomic profiling of neurological changes in valproic acid-treated mice.** *Journal of Proteome Research*, 12(5):2116-2127, 2013
- [qPSP] Goh et al. **Quantitative proteomics signature profiling based on network contextualization.** *Biology Direct*, 10:71, 2015
- [PFSNET] Goh & Wong. **Evaluating feature-selection stability in next-generation proteomics.** *Journal of Bioinformatics and Computational Biology*, 14(5):1650029, 2016
- [ESSNET] Goh & Wong. **Advancing clinical proteomics via analysis based on biological complexes: A tale of five paradigms.** *Journal of Proteome Research*, 15(9):3167-3179, 2016
- [PROTREC] Goh & Wong. **Recovering missing proteins based on biological complexes.** In preparation