Algorithms in Bioinformatics: A Practical Introduction

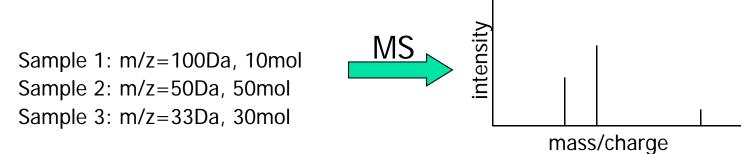
Peptide Sequencing

What is Peptide Sequencing?

- High-throughput Protein Sequencing is to deduce the amino acid sequence of a protein. It is still very difficult.
- Currently, research focus on Peptide Sequencing, that is, getting the amino acid sequence of a short fragment of a protein (of length ≈ 10).

Enabling technology: Mass Spectrometry

- Idea for deducing the peptide sequence: Mass!
- Mass Spectrometry is a machine which can separate and measure samples with different mass/charge ratio.
- Example:



Dalton(Da) is a mass unit. E.g. H is of mass 1Da

History

- Peptide sequencing is discovered by Pehr Edman (1949) and Frederick Sanger (1955).
- In 1966, Biemann et al successfully sequenced a peptide using a mass spectrometer machine.
- During 1980s, sequencing using mass spectrometry becomes popular.

Agenda

- Biological Background
- De Novo Peptide Sequencing
 - PEAK
 - Spectrum graph
- Protein Database Searching Problem
 - SEQUEST

Amino acid residue mass

Α	71.08	M	131.19
С	103.14	Ν	114.1
D	115.09	P	97.12
Е	129.12	Q	128.13
F	147.18	R	156.19
G	57.05	S	87.08
H	137.14	т	101.1
I	113.16	V	99.13
K	128.17	W	186.21
L	113.16	Y	163.18

- Amino acid residue
 amino acid losing
 a water
- I and L have the same mass
- Smallest mass is G (57.05 Da)
- Largest mass is W (186.21 Da)

Mass Spectrometry can separate different peptides

- Previous table shows that most of the amino acids have different masses.
- Hence, with high chance, different peptides have different masses.
- The mass given by a mass spectrometer has a maximum error ±0.5Da. It can separate most of the peptides.

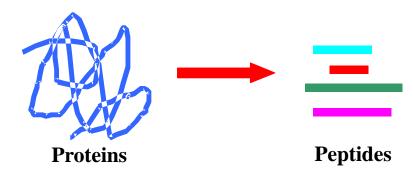
Protein identification process (LC/MS/MS)

Input: a protein sample

- A. Biology part:
 - 1. Digest the protein into a set of peptides
 - 2. By HPLC+Mass Spectrometer, separate the peptides.
 - 3. Select a particular peptide
 - 4. Fragment the selected peptide
 - 5. Get the tandem mass (MS/MS) spectrum of the selected peptide
- B. Computing part:
 - De Novo Sequencing
 - Protein Database Search

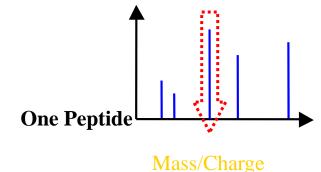
Digest a protein into peptides

- By an enzyme, digest a protein into short peptides.
- If we digest a protein using trypsin,
 - it digests the protein at K or R that are not followed by P.
 - After digestion, we will get a set of peptides end with K or R!
- E.g. ACCHCKCCVRPPCRCA → ACCHCK, CCVRPPCR



Selecting a particular peptide

- HPLC stands for High Performance Liquid Chromatograph. It can separate a set of peptides in a high pressure liquid chromatography
- After HPLC, the mixture of peptides are analyzed by MS.
 - Then, we get the MS spectrum



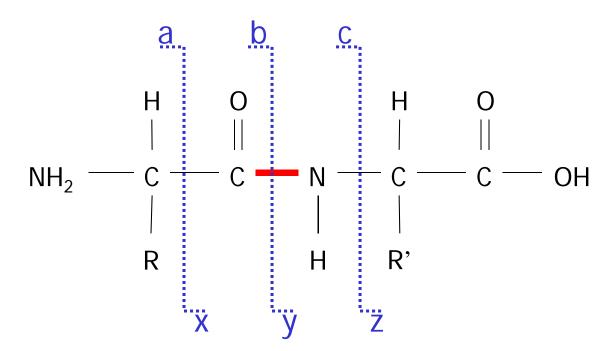
• The peptide of a particular mass is selected.

Fragmentation of peptide (I)

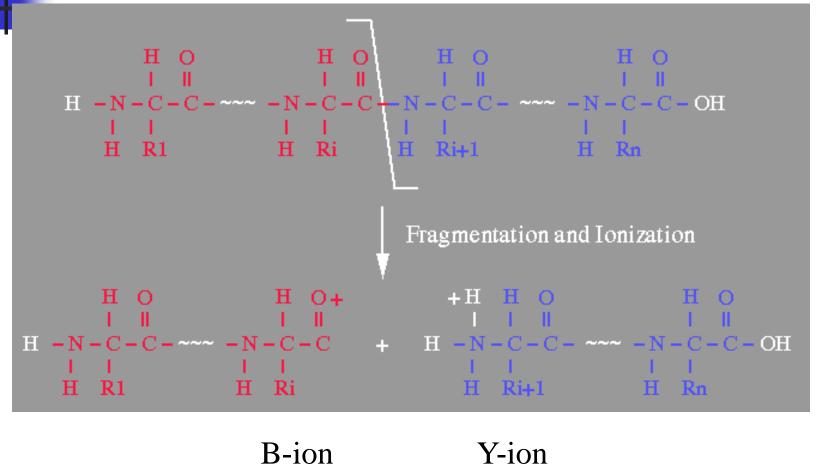
- Fragmentation tries to break the selected peptide at all positions in the peptide backbond.
- Usually, fragmentation is by Collision Induced Dissociation (CID).
 - The peptide is passed into the collision cell (which has been pressurized with argon [inert gas]).
 - Collision between peptide and argon break the peptide.
- Each peptide is usually fragmented into 2 pieces.
 - prefix fragment and suffix fragment (either one fragment will be charged but not both)

Fragmentation of peptide (II)

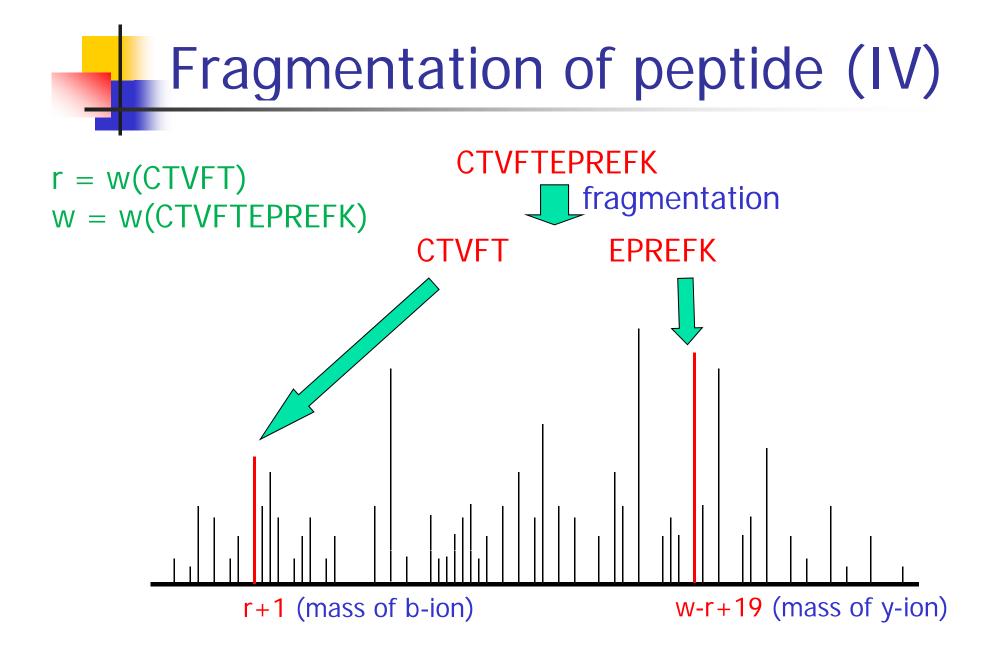
- Most often, the peptide is broken at C-C, C-N, N-C bonds.
 - Resulting a-ions, b-ions, c-ions, x-ions, y-ions, and z-ions.
 - Based on experiment,
 - The intensity of y-ions > that of b-ions
 - The intensities of other ions are even smaller



Fragmentation of peptide (III)



Complementary: Mass(B-ion)+Mass(Y-ion) = Mass(peptide)+4H+O



Mass of the ions (I)

- Let A be the set of amino acid. For every a∈A, w(a)
 = mass of its residue
- Let $P = a_1 a_2 \dots a_k$ be a peptide.

• $w(P) = \Sigma_{1 \le j \le k} w(a_j).$

- Actual mass of the peptide with sequence P is
 - w(P)+18 (since it has an extra H₂O)
- Mass of b-ion of the first i amino acids is

• $b_i = 1 + w(a_1a_2...a_i)$

Mass of y-ion of the last i amino acids is

• $y_i = 19 + w(a_i...a_k)$

• Note: $b_i + y_{i+1} = 20 + w(P)$

Mass of the ions (II)

- E.g. P=SAG
 - w(P) = w(S) + w(A) + w(G) = 215.21
 - Actual mass of P = w(P) + 18 = 233.21

•
$$y_1 = w(SAG) + 19 = 234.21$$

• $y_2 = w(AG) + 19 = 147.13$

•
$$y_3 = w(G) + 19 = 76.05$$

• $b_1 = w(S) + 1 = 88.08$

•
$$b_2 = w(SA)$$

• $b_3 = w(SAG) + 1 = 216.21$

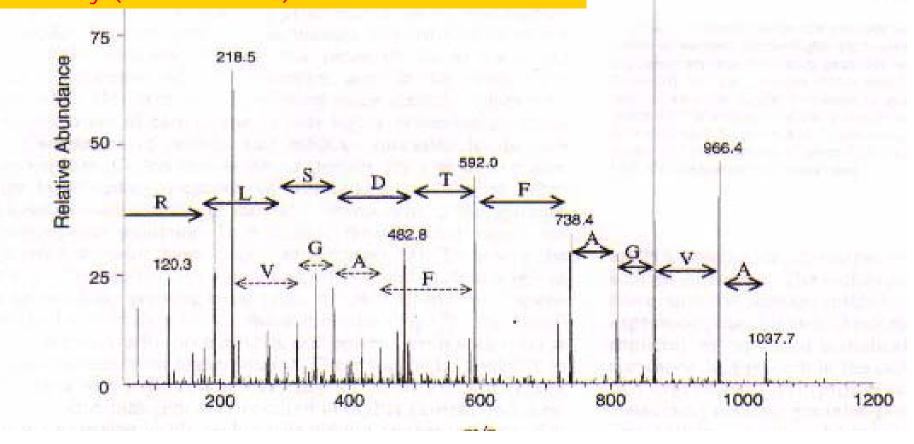
Other ion types

- Apart from a-ion, b-ion, c-ion, x-ion, y-ion, and z-ion, we also have variations with additional loss of
 - a water molecule
 - an ammonia molecule
 - a water and an ammonia molecule
 - Two water molecules

• E.g. $y-H_2O$, $y-NH_3$, $y-H_2O-H_2O$, $y-H_2O-NH_3$

Tandem Mass Spectrum (MS/MS Spectrum)

An MS/MS spectrum is represented as $M = \{(x_i, h_i) | 1 \le i \le n\}$ where x_i is the m/z for the i-th peak and h_i is its intensity (or abundance)



FAVGAFTDSLR

867.1

Computational problems

- There are three computational problems:
 - 1. De novo peptide sequencing
 - 2. Peptide Identification
 - 3. Identification of PTM (Post-translational modification)
- We will discuss problems 1 and 2.

De Novo Peptide Sequencing Problem

Input:

- A MS/MS spectrum M; and
- the total mass wt of the peptide
- An error bound δ (default δ =0.5)
- Output:
 - The peptide sequence

Assumption of the spectrum

- We assume all the ions are singly charged.
- In fact, in a MS/MS experiment,
 - an ion can be charged with different charges.
- Fortunately,
 - if a spectrum has peaks corresponding to multiply charged ions, there exists standard method to convert those peaks to their singly charged equivalents.

Simple scoring scheme

• Consider a peptide $P=a_1a_2...a_k$

- Recall that y-ions are expected to have the highest intensities.
- If M is a spectrum for P, we can find peaks for m/z = y_i for i=1,2,...,k
- So, we define the score function score(M,P) = $\Sigma{h|(x,h)\in M, |x-y_i|\leq \delta \text{ for } i=1,2,...,k}$

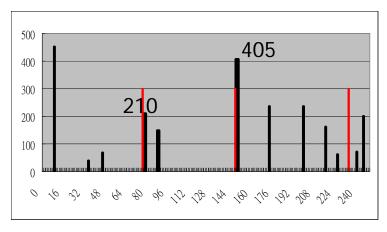
Simple scoring scheme example

• E.g. P=SAG

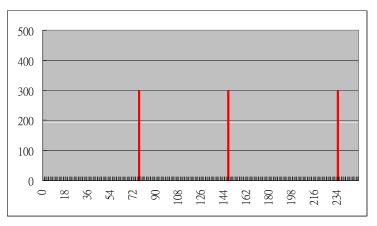
- $y_1 = 57.05 + 71.08 + 87.08 + 19 = 234.21$
- $y_2 = 57.05 + 71.08 + 19 = 147.13$

•
$$y_3 = 57.05 + 19 = 76.05$$

• Score(M,P) = 210+405 = 615



Black peaks: real peaks



Red peaks: artificial y-ions

Refined problem

Input:

- A MS/MS spectrum M
- The total mass wt of the peptide
- An error bound δ
- Output:
 - A peptide P such that wt-δ≤w(P)≤wt+δ which maximizes score(M,P).

Brute-force solution

- For every possible peptide P such that $|w(P)-wt| \le \delta$,
 - Compute score(M,P)
- Report the peptide P such that $|w(P)-wt| \le \delta$ which maximizes score(M,P)!
- Exponential time! Very slow!
- Can we solve the problem faster?
 - Yes! By dynamic programming.

Idea of the dynamic programming

- Try to identify the residues one by one from right to left.
- Let $f_M(r) = \Sigma \{ h \mid (x,h) \in M \text{ and } |x-r| \le \delta \}.$
 - $f_M(r)$ is the sum of all peaks in M whose mass is close to r.
- Observation:
 - score(M, $a_1a_2...a_k$) = score(M, $a_1a_2...a_{k-1}$)+f_M(w($a_1a_2...a_k$)+19)

Simple dynamic programming solution

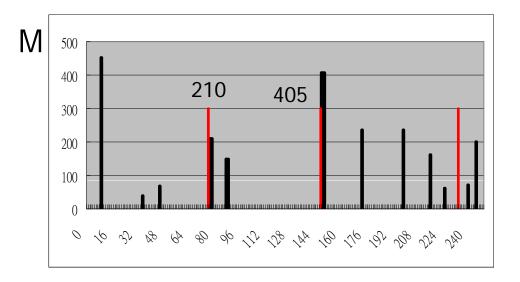
- Let V(r) be the maximum score(M,P) among all possible P such that w(P)=r.
- Our aim is to find max_{|r-wt|≤δ}V(r). Then, by back-tracking, we can recover the peptide.

We have

- V(0)=0.
- $V(r) = \max_{a \in A} \{ V(r-w(a)) + f_M(r+19) \}.$

Example

Recall V(0)=0. $V(r) = \max_{a \in A} \{ V(r-w(a)) + f_M(r+19) \}.$ E.g. $V(147.13) = \max \begin{cases} V(76.05) + 450 (due to A) \\ V(43.99) + 450 (due to C) \\ ... \end{cases}$





Algorithm Max_Y_Ion

Require: The mass spectrum M and a weight W

Ensure: A peptide P of mass between $W - \delta$ and $W + \delta$ which maximizes $score_Y(M, P)$.

1: Set
$$V(r) = 0$$
 for $r < 0$

2: for
$$r = 0$$
 to $W + \delta$ do

3:
$$V(r) = \max_{a \in A} \{ V(r - w(a)) + f_M(r + 19) \}$$

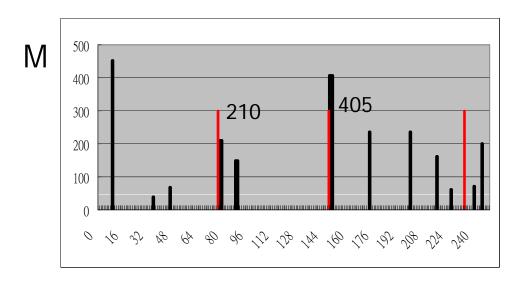
4: end for

5:
$$r' = \arg \max_{W - \delta \le r \le W + \delta} V(r)$$

6: Through back-tracing, we find the peptide P of mass r' which maximizes $score_Y(M,P)$

Example

- Given the spectrum M and wt=215.21.
 - V(76.05) = V(0) + 210 = 210 (due to G)
 - V(147.13) = V(76.05) + 450 = 615 (due to A)
 - V(234.21) = V(147.13) + 0 = 615 (due to S)
- By backtracking, we recover SAG!



Time analysis

- We need to fill-in the V table with wt entries.
- Each entry can be computed in O(|A|) time.
- So, total time complexity is O(|A|wt) time.

Can we use more information other than y-ions?

Yes. We can also use information from b-ions.

Better scoring scheme

Consider a peptide P=a₁a₂...a_k

- If M is a spectrum for P, we can find peaks for m/z = y_i or m/z = b_i for i=1,2,...,k
- So, we redefine the score function score(M,P) as Σ{h|(x,h)∈M, |x-y_i|≤δ or |x-b_i|≤δ for i=1,2,...,k}

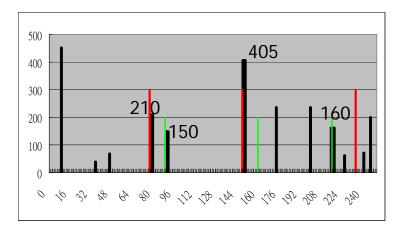
Better scoring scheme example

- E.g. P=SAG
 - $y_1 = 57.05 + 71.08 + 87.08 + 19 = 234.21$
 - $y_2 = 57.05 + 71.08 + 19 = 147.13$
 - $y_3 = 57.05 + 19 = 76.05$
 - $b_1 = 87.08 + 1 = 88.08$
 - $b_2 = 87.08 + 71.08 + 1 = 159.16$
 - $b_3 = 87.08 + 71.08 + 57.05 + 1 = 216.21$

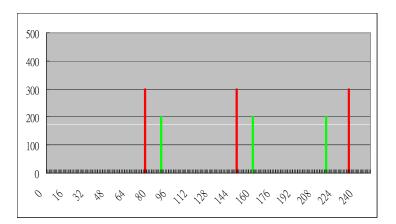


= 210 + 405 + 150 + 160

= 925



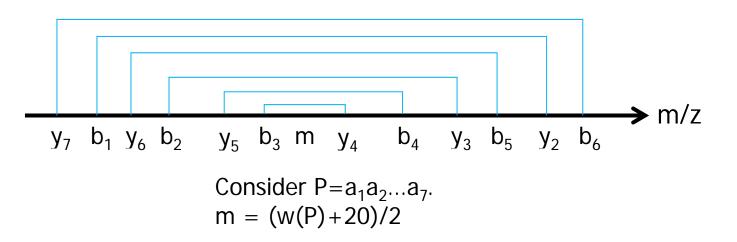
Black peaks: real peaks Green peaks: artificial b-ions



Red peaks: artificial y-ions

Observations

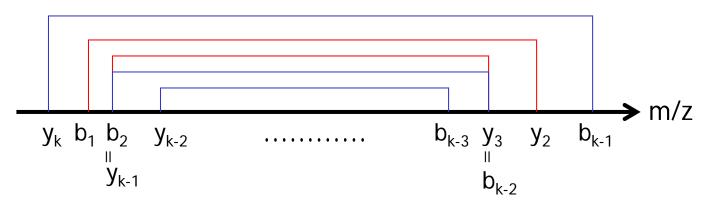
- Suppose $P=a_1a_2...a_k$.
- b_i is strictly increasing while y_j is strictly decreasing.
 - Proof: For any peptide Q and amino acid a, w(Qa), w(aQ) > w(Q).
 - Hence, $b_{i+1}-b_i$, $y_j-y_{j+1} \ge \min_{a \in A} w(a) = 57.05 > 0$
- 2. Note that $b_i + y_{i+1} = w(P) + 20$.
 - Hence, we have (b_i, y_{i+1}) , for all i=1,2,...,k, form a set of nested regions.
 - For the adjacent nested intervals, the mass different is at most max_{a∈A}w(a) = 186.21.



Can we solve the problem using previous DP?

No!

- The reason is that, for some masses y_i and b_j, their masses may be very close and correspond to the same peak (x, h)∈M.
- In this case, the previous DP will sum the same peaks two times.



Observation (II)

- Note that the outermost ℓ intervals are formed by breaking the prefix $a_1...a_i$ and the suffix $a_j...a_k$, where $i+(k-j+1)=\ell$.
- Let score'($M, a_1...a_i, a_j...a_k$) be
 - the sum of the intensities of all b-ion and y-ion peaks formed by breaking the peptide P between a_x and a_{x+1} for x∈{1,...,i}∪{j-1...,k-1}.
- Let f_M(r,s) be the sum of all peaks in M which are close to r and wt+20-r but not close to s and wt+20-s. [used to avoid double counting!]
- We have

 $score'(M, a_1 \dots a_i, a_j \dots a_k) = \begin{cases} score'(M, a_1 \dots a_{i-1}, a_j \dots a_k) + f_M(b_i, y_j) \text{ if } b_i \ge y_j \\ score'(M, a_1 \dots a_i, a_{j+1} \dots a_k) + f_M(y_j, b_i) \text{ otherwise} \end{cases}$

Solution (a more complicated dynamic programming)

• Let \hat{a} be $\max_{a \in A} w(a) = 186.21$.

■ For every $|r-s| \le \hat{a}$, let V(r, s) be the maximum score'(M,P₁,P₂) among all possible P₁ and P₂ where w(P₁)=r and w(P₂)=s.

$$V(r,s) = \max \begin{cases} \max_{a \in A} \{ V(r - w(a), s) + f_M(r + 1, s + 19) \}, \ r \ge s \\ \max_{a \in A} \{ V(r, s - w(a)) + f_M(s + 19, r + 1) \}, \ r < s \end{cases}$$

with base case $V(r,s) = 0 (r \le 0, s \le 0)$.

Solution (a more complicated dynamic programming)

- Aim: Find the best V(r,s) such that wt+20=r+s+w(a) for some a∈A.
 - Then, by back-tracking, we can recover the peptide.

Algorithm Sandwich

Require: A mass spectrum M, a weight W, and an error bound δ **Ensure:** A peptide P such that score(M, P) is maximized and |w(P) - $|W| < \delta$ 1: Let $\hat{a} = \max_{a \in A} w(a)$ 2: Initialize all $V(r,s) = -\infty$; Let V(0,0) = 03: for r = 1 to $(W/2 + \hat{a})$ do for $s = r - \hat{a}$ to min $\{r + \hat{a}, W - r\}$ do 4: for $a \in A$ such that r + s + w(a) < W do 5: if r < s then 6: $V(r,s) = \max\{V(r,s), V(r-w(a),s) + f_M(r+1,s+19)\}$ 7: else 8: $V(r,s) = \max\{V(r,s), V(r,s-w(a)) + f_M(s+19,r+1)\}$ 9: end if 10: end for 11: end for 12:13: end for 14: Identify the best V(r,s) among all r, s, a satisfying $|r-s| < \hat{a}$ and $|r+s+w(a)-W| < \delta$. Through back-tracing, we can recover a peptide P = P'aP'' where w(P') = r and w(P'') = s.

Time complexity

- We need to fill-in V(r,s) for all $|r-s| \le \hat{a}$.
- So, we need to fill-in wt·â entries.
- Each can be filled-in using O(|A|) time.
- The time complexity is O(wt·â·|A|) time.

Spectrum Graph approach

Another method to recover the peptide is based on spectrum graph, which is defined as follows. Generating vertices in the spectrum graph

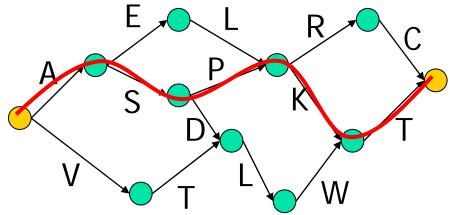
- For each mass r in the spectrum M,
 - We generate two vertices of masses r and wt-r.
- We also include 2 additional vertices:
 - starting vertex with mass = 0 and
 - ending vertex with mass = wt.

Generating edges in the spectrum graph

- For every pair of mass r and s,
 - If r-s equals the mass of an amino acid A,
 - we connect x and y with an edge of label A.
- Since there may be some missing peaks in the spectrum,
 - If r-s equals the total mass of two amino acids A₁A₂,
 - we connect x and y with an edge of label A_1A_2 .
 - If r-s equals the total mass of three amino acids A₁A₂A₃,
 - we connect x and y with an edge of label $A_1A_2A_3$.

Meaning of a path in the graph

- Every path from start to end corresponds to a possible peptide in the spectrum
- However, there are many possible paths?



Weight of the edges

- Observe that a vertex has higher probability to be real if all ion types are available.
- Hence, we can assign a score depending on whether some ion types are missing.
- Then, this is a problem of finding the heaviest path, which can be solved in polynomial time.

Weighting function for Sherenga

- Assume noise is produced uniformly and randomly with probability q_R.
- Assume q_b is the probability that the b-ion peak exists in M given the bion appears in the theoretical spectrum.
- Similarly, assume q_y is the probability that the y-ion peak exists in M given the y-ion appears in the theoretical spectrum.
- The weight of every vertex with mass v is defined as the sum of score_b(v) and score_v(v), where

$$score_b(v) = \begin{cases} \log \frac{q_b}{q_R} & \text{if } v + 1 \text{ exists in } M \\ \log \frac{1-q_b}{1-q_R} & \text{otherwise} \end{cases}$$

$$score_{y}(v) = \begin{cases} \log \frac{q_{y}}{q_{R}} & \text{if } W - v + 19 \text{ exists in } M \\ \log \frac{1 - q_{y}}{1 - q_{R}} & \text{otherwise} \end{cases}$$

Protein Database searching Problem

- Input:
 - a database of proteins (DB)
 - a raw MS/MS spectrum (M)
 - The mass wt of the peptide corresponding to M

• Output:

- A protein whose peptide is expected to have mass wt and a MS/MS spectrum similar to M.
- This lecture presents a solution called SEQUEST (Eng et al, 1994)

SEQUEST

- Step 1: Reduction of the tandem mass spectrometry data
 - To avoid noise, only 200 most abundant signals of the raw spectrum are used.
 - Also, the total signals of the 200 signals are renormalized to 100.
- Step 2: Search the protein database DB to find all peptides such that each peptide P has mass within (wt±1)Da

SEQUEST

 Step 3: Rank the top 500 fit sequences by a specific scoring function.

SEQUEST

- Step 4: Compare the spectral similarity. Use cross-correlation analysis to generate the final score and rank the sequences.
- The abundance of ions in the hypothetic spectrum: 50 (b-ion, y-ion), 25 (mass/charge within ±1 from b or y), or 10 (a-ion)

Conclusion

- This lecture presents two De Novo Peptide Sequencing algorithms.
- We also present the protein database searching algorithm SEQUEST.
- There are many other problems in this area. For example,
 - Identifying peptide modifications

References

- J. K. Eng, A. L. McCormack, J. R. Yates. "An approach to correlate tandem mass spectral data of peptides with amino acid sequences in a protein database". J. Am. Soc. Mass Spectrom, 5:976-989, 1994.
- B. Ma, K. Zhang, C. Liang. "An Effective Algorithm for the Peptide De Novo Sequencing from MS/MS Spectrum". CPM, 266-277, 2003.
- V. Dancik, T. A. Addona, K. R. Clauser, J. E. Vath, P. A. Pevzner. De novo peptide sequencing via tandem mass spectrometry. Journal of Computational Biology, 6:327-342, 1999.