Gene expression analysis: Some lessons for statistical hypothesis testing

WONG Limsoon



Plan, 12/1/2026

Statistical hypothesis testing

Onus of proof

Anna Karenina Principle

Null hypothesis & null distribution

Getting them right

Gene expression profiling

Differentially expressed genes (DEG) selection

Poor DEG selection replicability

Addressing the replicability crisis

Statistical hypothesis testing

Formulate null hypothesis H0 and alternate hypothesis H1

Devise a test statistic, $t(\cdot)$

Evaluate t(S) on a sample S

Compare t(S) to the null distribution

This does not mean we accept H0!

If significant, reject H0; otherwise, reject H1

Null distribution is the distribution of t(S0) where S0 ranges over the set of null samples for which H0 holds

Onus of proof: Reject H1 ≠ accept H0

"... a p-value is large doesn't mean that the null hypothesis is true. All a hypothesis test does is measure the strength of evidence against the null hypothesis. That is, we assume the null hypothesis is true until we have enough evidence to reject. Crucially, we never actually claim that the null hypothesis is true - it is just an assumption!"

A pharmaceutical research team constructs a significance test:

HO – Side effects of new drug X are same as standard drug Y

H1 - Side effects of new drug X are different from standard drug Y

Would you be happy to use the new drug based on a large statistically insignificant p-value?

Anna Karenina Principle

There are many ways to violate the null hypothesis but only one way that is truly pertinent to the outcome of interest

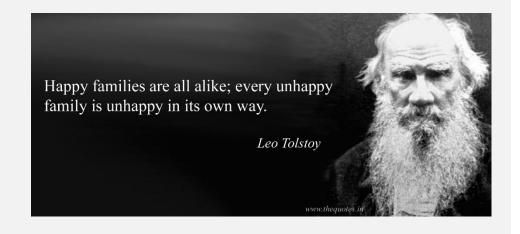
Sample is biased

Null distribution used is inappropriate

Null hypothesis incorrectly stated

Inappropriate expt design

And so on



Exercise

SNP rs123 is good biomarker for a disease:

If rs123 is AA or GG, low risk for disease

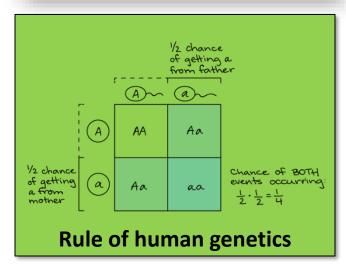
If rs123 is AG, high risk for disease

Group								
SNP	Genotypes	Cont	rols [n(%)]	Cases	s [n(%)]	χ ²	P value	
rs123	AA	1	0.9%	0	0.0%		4.78E-21 ^b	
	AG	38	35.2%	79	97.5%			
	GG	69	63.9%	2	2.5%			
Abbreviation: SNP, single nucleotide polymorphism.								

Straightforward χ 2 test. Anything wrong?

Calculations

	Group							
SNP	Genotypes	Cont	rols [n(%)]	Case	s [n(%)]	χ²	<i>P</i> value	
rs1	AA	1	0.9%	0	0.0%		4.78E-21 ^b	
23	AG	38	35.2%	79	97.5%			
	GG	69	63.9%	2	2.5%			



Controls + cases = 189

 \Rightarrow Population is 117 / 189 = 62% AG

$$P(AA) = ...$$



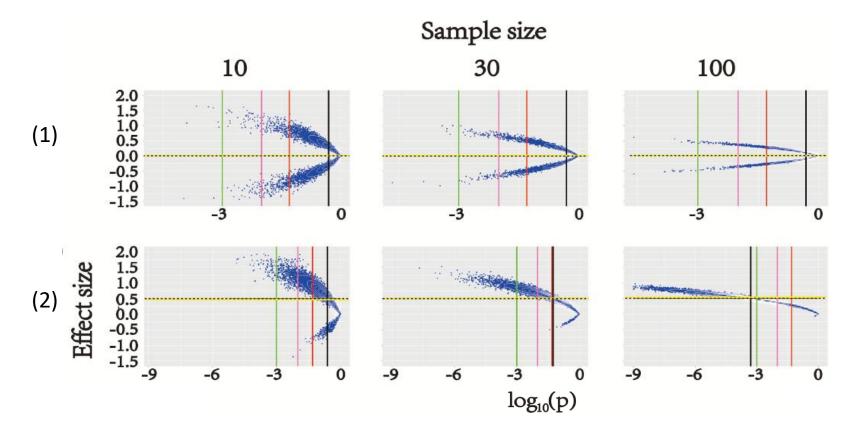
Sample may be ...

What do you think?



Sampling bias happens often

	Dis	tribution		Mean	Standard o	deviation	Sample size		
Scenario	A	В	A	В	A	В			
(1)	Normal	Normal	0	0	1	1	10	30	100
(2)	Normal	Normal	0	0.5	1	1	10	30	100



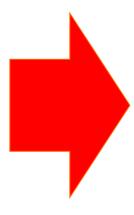
Careless null hypothesis

H₀

rs123 alleles are identically distributed in the two samples

Assumption

Distributions of rs123 alleles in the two samples are resp. identical to the two populations



Apparent H1

rs123 alleles are differently distributed in the two populations

"Actual" H1

rs123 alleles are differently distributed in the two populations OR

Distribution of rs123 alleles in the two samples are not identical to the two populations

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Exercise

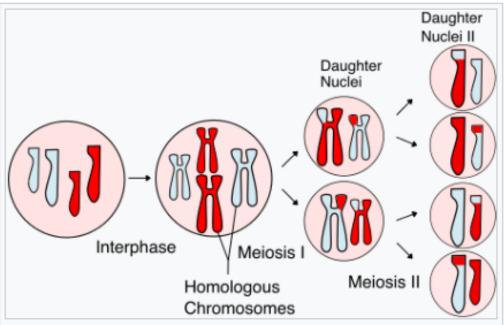
Suppose distributions of rs123 alleles in the two samples are identical to the corresponding populations and the test is significant

Can we say rs123 mutation causes the disease?

Wong Limsoon, BS6213, Jan 2026

Meiosis

Watch this video together, https://youtu.be/BlnUNmfGn71



In meiosis, the chromosomes duplicate (during interphase) and homologous chromosomes exchange genetic information (chromosomal crossover) during the first division, called meiosis I. The daughter cells divide again in meiosis II, splitting up sister chromatids to form haploid gametes. Two gametes fuse during fertilization, forming a diploid cell (zygote) with a complete set of paired chromosomes.

Image credit: Wikipedia

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When two genes are close together, this is what happens during meiosis

Watch this video together, https://youtu.be/ZeATszO-6e0

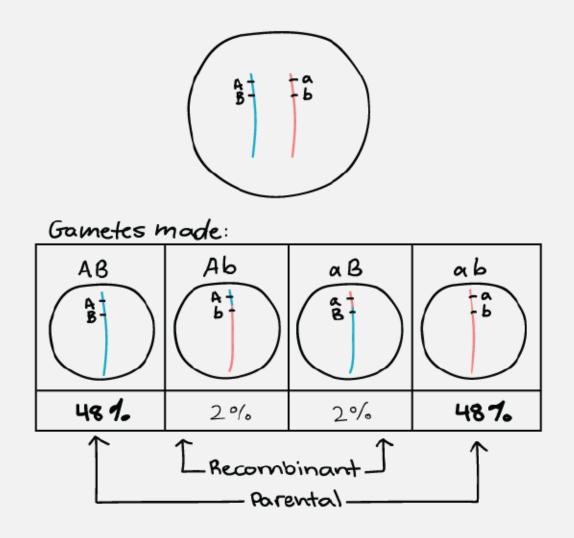
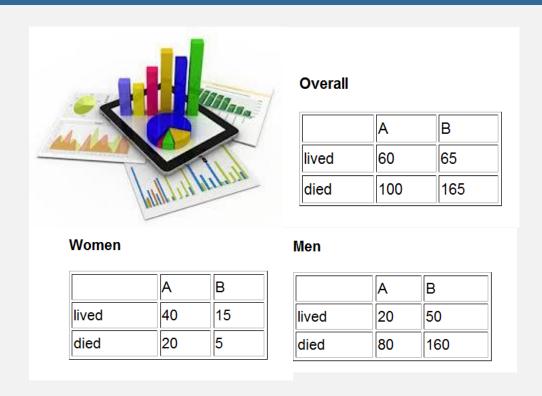


Image credit: Khan Academy

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Exercise



Treatment A is better



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Treatment B is better

What is happening here?

A/B sample not ...

What do you think?



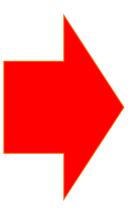
Careless null hypothesis

H0

Treatment effects are identically distributed in the two samples

Assumption

All other factors are equalized in the two samples



Apparent H1

Treatment effects are differently distributed in the two populations

"Actual" H1

Treatment effects are differently distributed in the two populations OR

Some other factors aren't equalized in the two samples

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Exercise

I asked ChatGPT how to test whether my income is atypically low

What do you think of ChatGPT's response?



ChatGPT

To test whether your income is atypically low given the mean income (X) and standard deviation (Y), you can use z-scores to determine how many standard deviations your income is away from the mean. This helps identify whether your income is significantly lower or higher than the average.

The z-score formula is:

$$z = \frac{\text{Your Income-Mean Income}}{\text{Standard Deviation}} = \frac{\text{Your Income-}X}{Y}$$

Once you calculate the z-score for your income using this formula, you can interpret its value:

- If the z-score is significantly less than -2 (approximately), your income might be considered atypically low.
- A z-score less than -2 indicates that your income is more than two standard deviations below the mean income, which is relatively rare in a normal distribution.

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A nation of unusually poor people, really?

What do you think



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Microarray

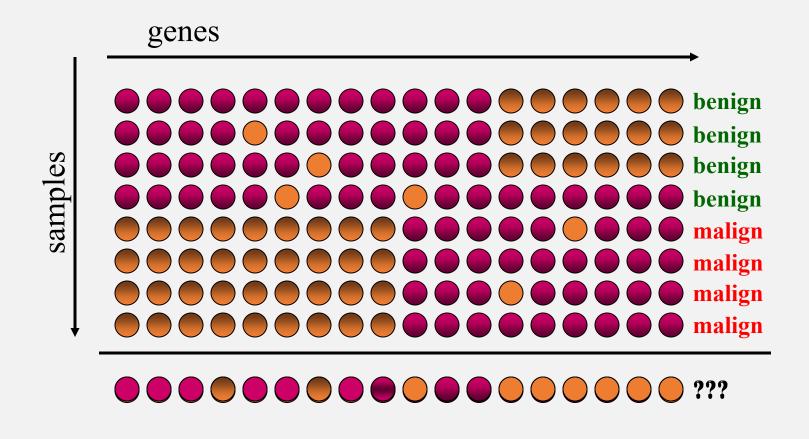
Contain large numbers of DNA molecules spotted on glass slides, nylon membranes, or silicon wafers

Detect what genes are being expressed or found in a cell of a tissue sample

Measure expression of thousands of genes simultaneously

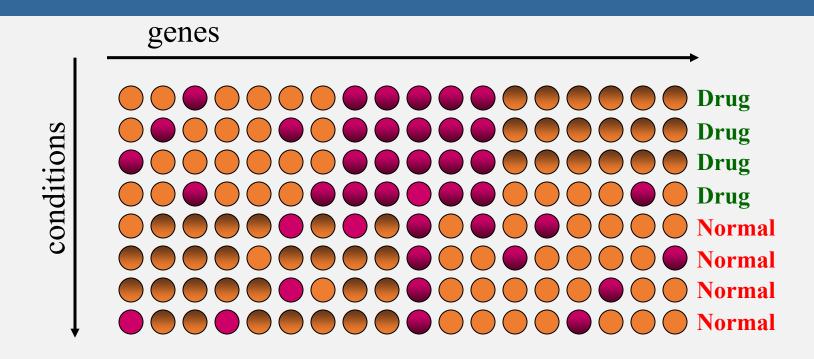
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Application: Disease subtype diagnosis



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Application: Drug-action inference



Which group of genes does the drug affect? Why?

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Diagnosis using microarrays & machine learning

Gene expression data collection

Gene selection using e.g., $\chi 2$

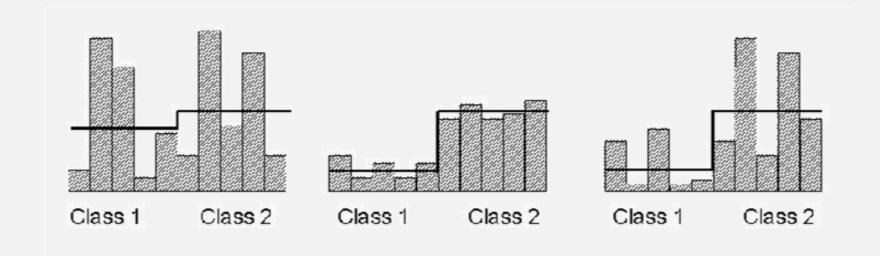
Classifier training

Classifier tuning (optional for some machine learning methods)

Apply classifier for diagnosis of future cases

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Gene selection basic idea



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Gene selection by $\chi 2$

The \mathcal{X}^2 value of a signal is defined as:

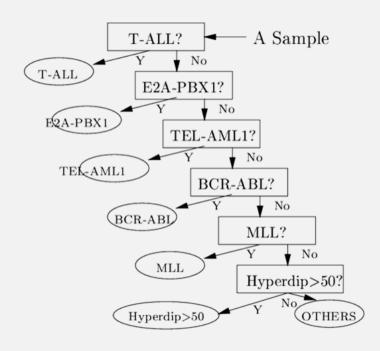
$$\mathcal{X}^{2} = \sum_{i=1}^{m} \sum_{j=1}^{k} \frac{(A_{ij} - E_{ij})^{2}}{E_{ij}},$$

where m is the number of intervals, k the number of classes, A_{ij} the number of samples in the ith interval, jth class, R_i the number of samples in the ith interval, C_j the number of samples in the jth class, N the total number of samples, and E_{ij} the expected frequency of A_{ij} ($E_{ij} = R_i * C_i/N$).

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Performance of various classifiers

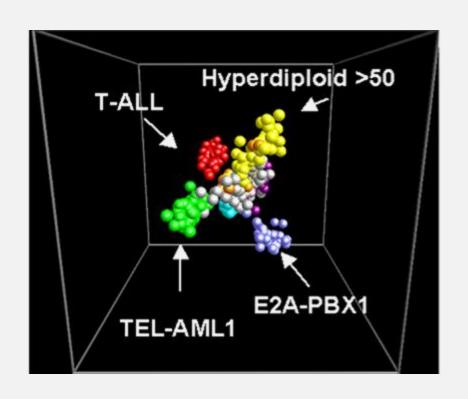
Testing Data	Error rate of different models				
	C4.5	SVM	NB	PCL	
T-ALL vs OTHERS1	0:1	0:0	0:0	0:0	
E2A-PBX1 vs OTHERS2	0:0	0:0	0:0	0:0	
TEL-AML1 vs OTHERS3	1:1	0:1	0:1	1:0	
BCR-ABL vs OTHERS4	2:0	3:0	1:4	2:0	
MLL vs OTHERS5	0:1	0:0	0:0	0:0	
Hyperdiploid>50 vs OTHERS	2:6	0:2	0:2	0:1	
Total Errors	14	6	8	4	

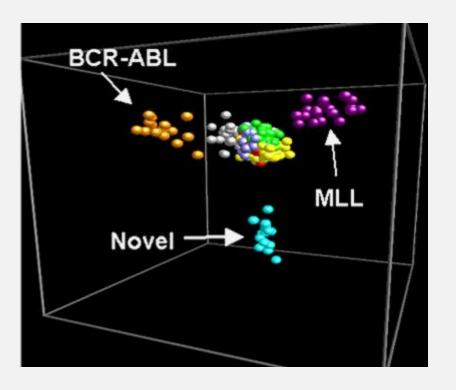


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Classifiers based 20 genes selected by $\chi 2$ at each level of the tree

Multidimensional scaling plot for subtype diagnosis





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Obtained by performing PCA on the 20 genes chosen for each level

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Poor replicability of gene selection

Low % of overlapping genes from diff expt

Prostate cancer

• Lapointe et al, 2004 vs Singh et al, 2002

Lung cancer

• Garber et al, 2001 vs Bhattacharjee et al, 2001

DMD

Haslett et al, 2002 vs Pescatori et al, 2007

Datasets	DEG	POG		
Prostate	Top 10	0.30		
Cancer	Top 50	0.14		
	Top100	0.15		
Lung	Top 10	0.00		
Cancer	Top 50	0.20		
	Top100	0.31		
D.11D	Top 10	0.20		
DMD	Top 50	0.42		
	Top100	0.54		

Individual genes

Suppose:

Each gene has 50% chance to be high You have 3 disease and 3 normal samples

How many genes on a microarray are expected to perfectly correlate to these samples?

Prob(gene is correlated) = $1/2^6$

of genes on array = 25,000

E(# of correlated genes) = 390

⇒ Many false positives; these cannot be eliminated based on pure statistics!

Group of genes

Suppose:

Each gene has 50% chance to be high You have 3 disease & 3 normal samples

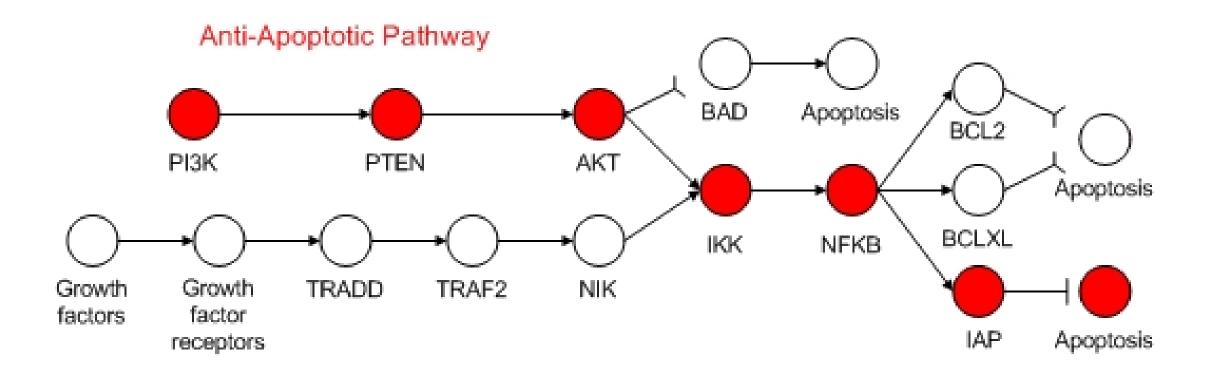
What's the chance for a group of 5 genes to perfectly correlate to these samples?

When only 1 group is considered, $(1/2^6)^5 << 1/2^6$ # of groups = $^{25000}C_5$ E(# of correlated groups) = $^{25000}C_5^*$ (1/2⁶)⁵ = 7.58 * 10 ¹⁰

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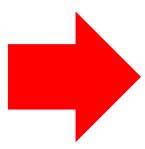
 \Rightarrow Even more false positives?

Perhaps no need to consider every group



Each disease phenotype has some underlying cause

There is some unifying biological theme for genes that are truly associated with a disease subtype



Uncertainty in selected genes reduced using biological processes

The unifying biological theme is basis for inferring underlying cause of disease subtype

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Group of Genes

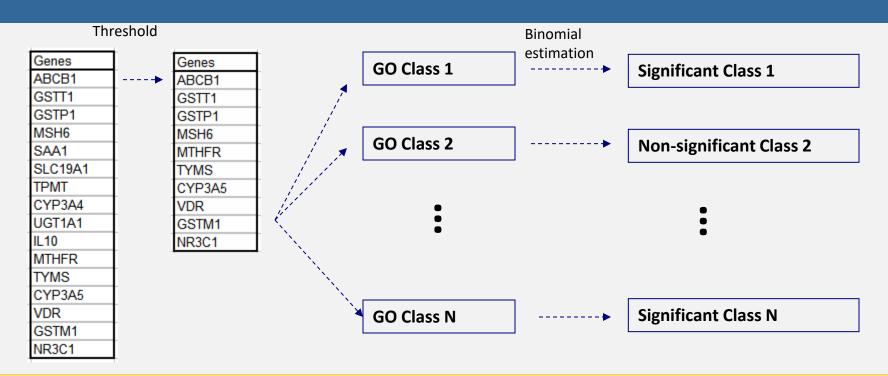
- Suppose
 - Each gene has 50% chance to be high
 - You have 3 disease and 3 normal samples
- What is the chance of a group of 5 genes being perfectly correlated to these samples?

- Prob(group of genes correlated) = $(1/2^6)^5$
 - Good, << 1/26
- # of groups = 25000 C₅
- ⇒ E(# of groups of genessis) correlated) = $\frac{25000}{5}$ C₅**
 (1/26)⁵ = 7.58*10¹⁰
- ⇒ Even more false positives?
- Perhaps no need to consider every group

of pathways = 1000

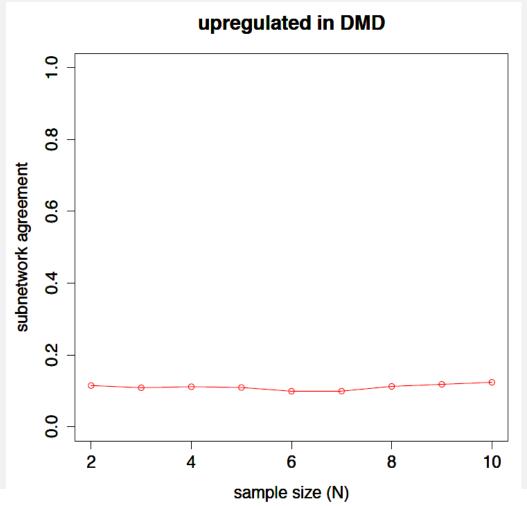
E(# of pathways correlated) = $1000 * (1/2^6)^5 = 9.3*10^{-7}$

Pathway overlap analysis via Onto-Express (aka ORA)



ORA tests whether a pathway is significant by intersecting the genes in the pathway with a predetermined list of DE genes (e.g., genes whose t-statistic meets the 5% significance threshold of t-test), and checking the significance of the size of the intersection using the hypergeometric test

The bewilderment persists... a crisis?



DMD gene expression data

- Pescatori et al., 2007
- Haslett et al., 2002

Pathway data

PathwayAPI, Soh et al., 2010

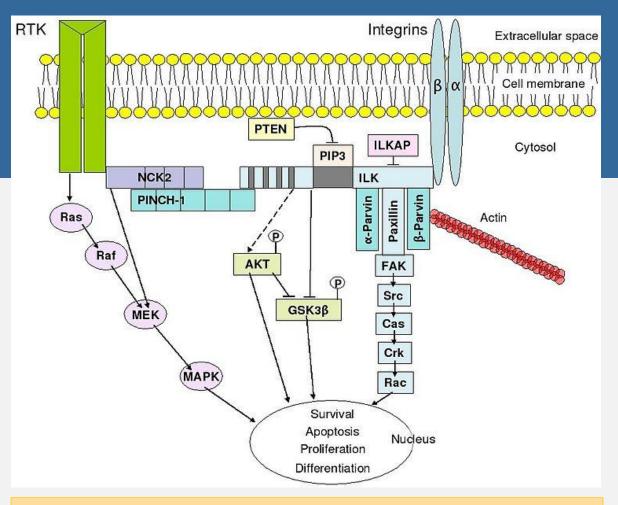
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Exercise

Why does ORA perform so poorly in selecting differentially expressed genes?

Issue #1 with ORA

What do you think?



A biological pathway is a chain of actions of molecules in cell leading to a change in cell

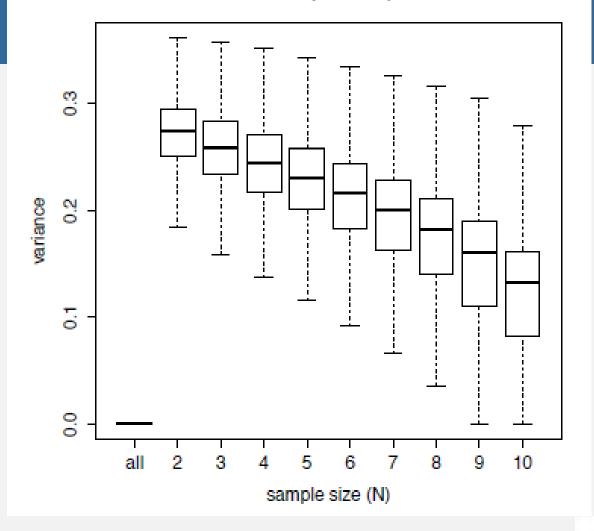
⇒ Behavour of genes in a pathway is more coordinated than random ones



Issue #2 with ORA

What do you think?

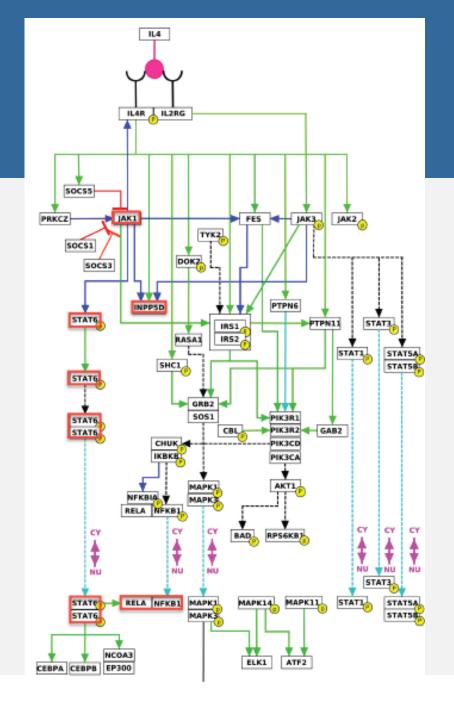
t-test p.value(s)





Issue #3 with ORA

What do you think?





Exercise

How to solve the issues identified in ORA?

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Take-home messages

Statistical hypothesis testing needs careful thought

Right null hypothesis

Right null distribution

Many nuances wrt confounding factors

Make effort to understand the domain

A little domain insight goes a really long way

References

Draghici et al., "Global functional profiling of gene expression", *Genomics*, 81(2):98-104, 2003

Lim et al., "A quantum leap in the reproducibility, precision, and sensitivity of gene expression profile analysis even when sample size is extremely small", *Journal of Bioinformatics and Computational Biology*, 13(4):1550018, 2015

Homework, due 17/1/2026

submit by email to dcswls@nus.edu.sg

Read this paper, focusing on how it proposed to test synthetic-lethal gene pairs:

Sriganesh et al., "Inferring synthetic lethal interactions from mutual exclusivity of genetic events in cancer", *Biology Direct*, 10:57, 2015

Read the review report (attached below) produced by ChatGPT discuss whether the proposed test is a good one.

Answer the following questions:

1/ Which parts of the ChatGPT report, if any, do you agree with?

2/ Which parts of the ChatGPT report, if any, do you disagree with? Why?

3/ Are there any major flaws that you think should be pointed out but are missed by ChatGPT?

Make 5-minutes presentation to class on 19/1/2026

Review Report

This manuscript proposes a computational method to infer synthetic-lethal (SL) interactions from tumour cohorts by identifying mutually exclusive patterns of genetic alteration. The central idea is that if gene A and gene B are each frequently altered but rarely co-altered, the missing double state reflects negative selection from lethality.

How the study tests for SL

1. Mutual-exclusivity test

For each gene pair, the authors tabulate the frequencies of A-only, B-only, and A+B alterations and apply a 1 – hypergeometric test to determine whether co-alterations occur significantly less often than expected under independence. Significant underrepresentation is taken as evidence of a synthetic-lethal relationship.

2. Alteration patterns considered

The method includes both loss–gain (e.g., BRCA2 loss with TLK2 amplification) and loss–loss (parallel DDR deficiencies) combinations. Treating these together is reasonable because SL may arise either from disrupting complementary pathways or from breaking regulatory feedback loops.

3. Validation with essentiality screens

The authors use genome-wide essentiality data from cell lines deficient in gene A. Candidate B genes are consistently more essential in A-defective lines than in proficient controls. This context-specific essentiality provides strong functional support for the SL interpretation.

4. Biological plausibility

Several predicted pairs match known interactions (e.g., PIK3CA-PTEN), and many candidate B genes correlate with poorer patient survival when overexpressed, consistent with tumour fitness roles.

Is their test a good one?

Strengths

- Directly leverages human tumour data.
- Statistical test appropriately detects underrepresented double events.
- Essentiality validation strengthens confidence in SL predictions.
- Context comparisons (e.g., PTEN^+/+ vs PTEN^-/-) support causality.

Limitations

- Mutual exclusivity can arise from non-SL causes (subtypes, pathway saturation).
- Limited power for rare alterations.
- Cell-line essentiality may not fully reflect tumour biology.

Overall assessment

Mutual exclusivity alone cannot prove SL, but paired with solid essentiality validation and plausible mechanisms, the approach is well justified and useful for prioritizing SL candidates in tumours.

Presentation & discussion on ...

Research Open access Published: 01 October 2015

Inferring synthetic lethal interactions from mutual exclusivity of genetic events in cancer

<u>Sriganesh Srihari, Jitin Singla, Limsoon Wong</u> № <u>& Mark A. Ragan</u>

Biology Direct 10, Article number: 57 (2015) Cite this article

