

For written notes on this lecture, please read Chapters 4 and 7 of *The Practical Bioinformatician*

CS2220: Introduction to Computational Biology

Lecture 4: Gene Feature Recognition

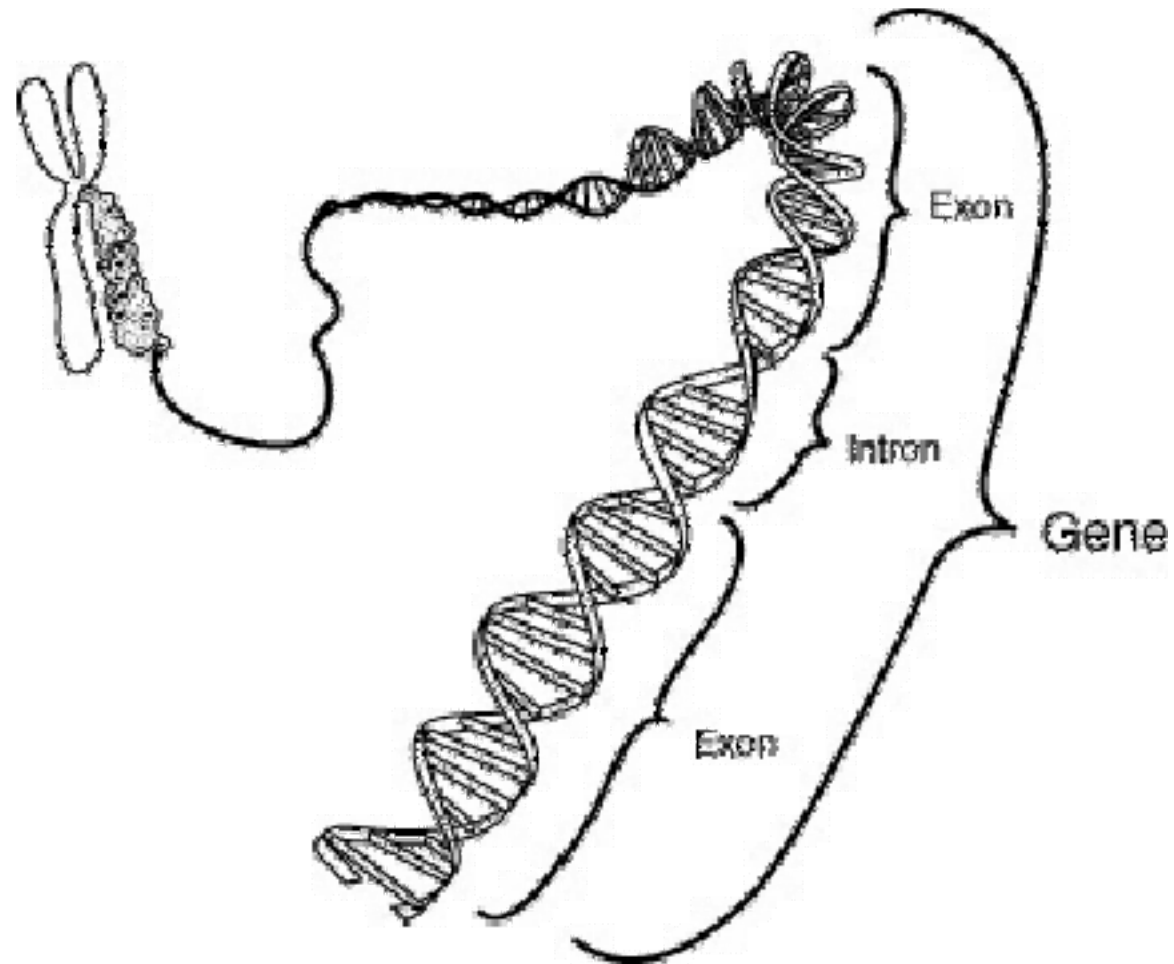
Limsoon Wong
2 February 2007



Central Dogma of Molecular Biology

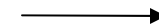
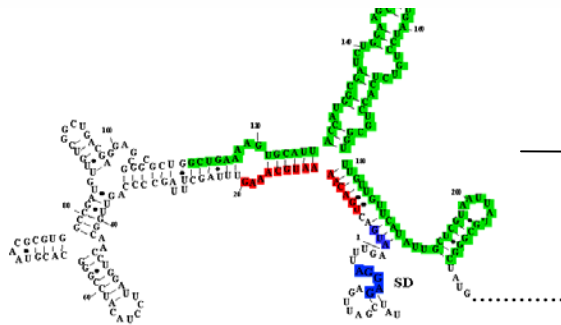
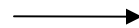
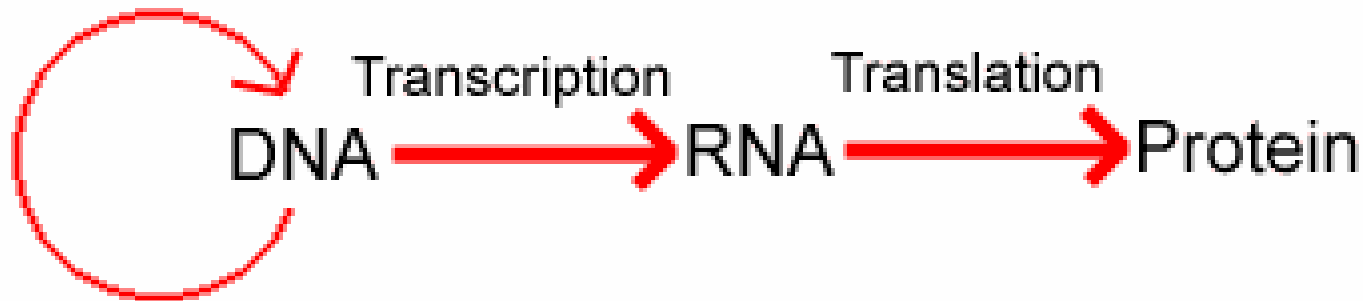


What is a gene?



Central Dogma

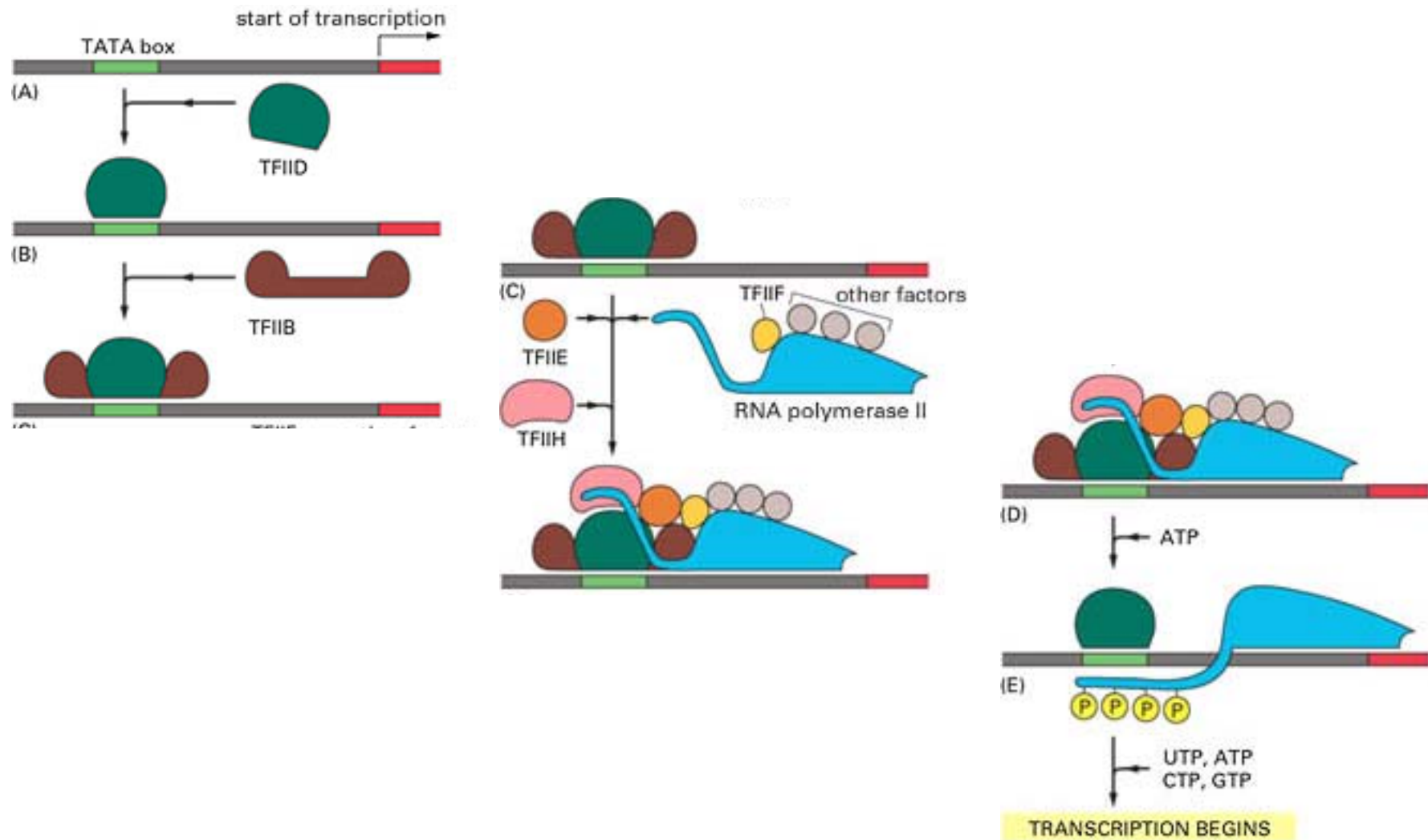
Replication



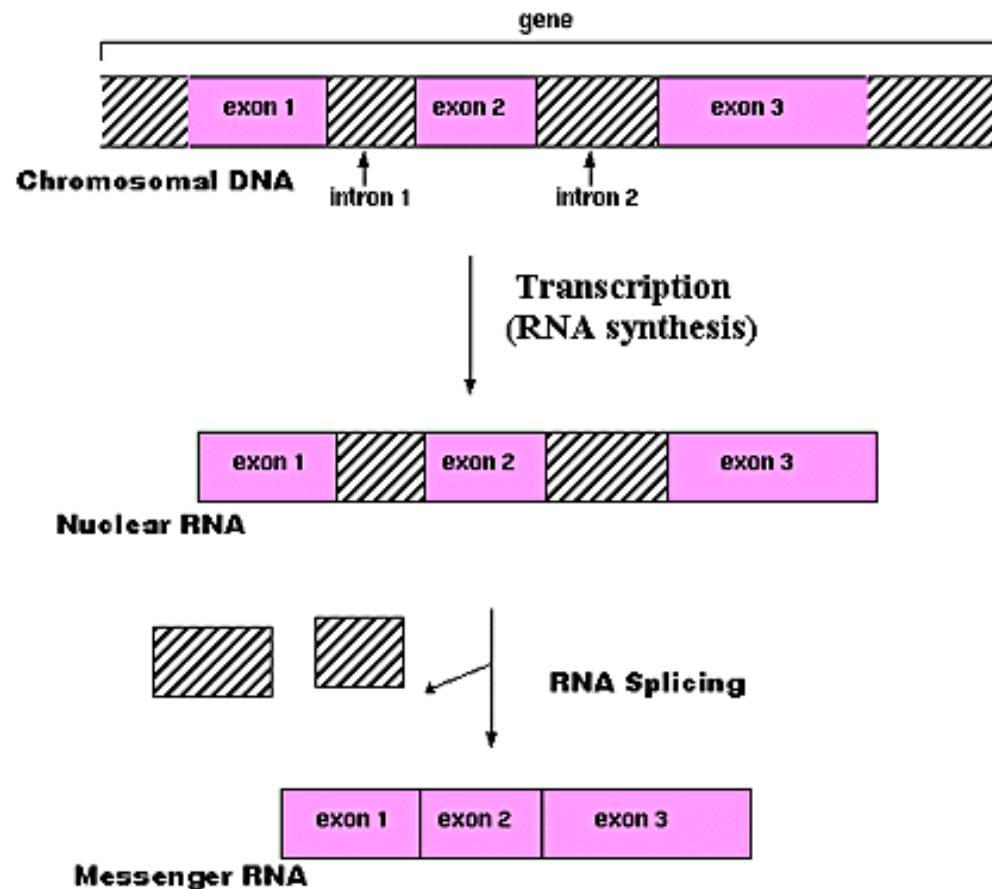
...AATGGTACCGATGACCTG...

...TRLRPLLALLALWP...

Transcription: DNA → nRNA

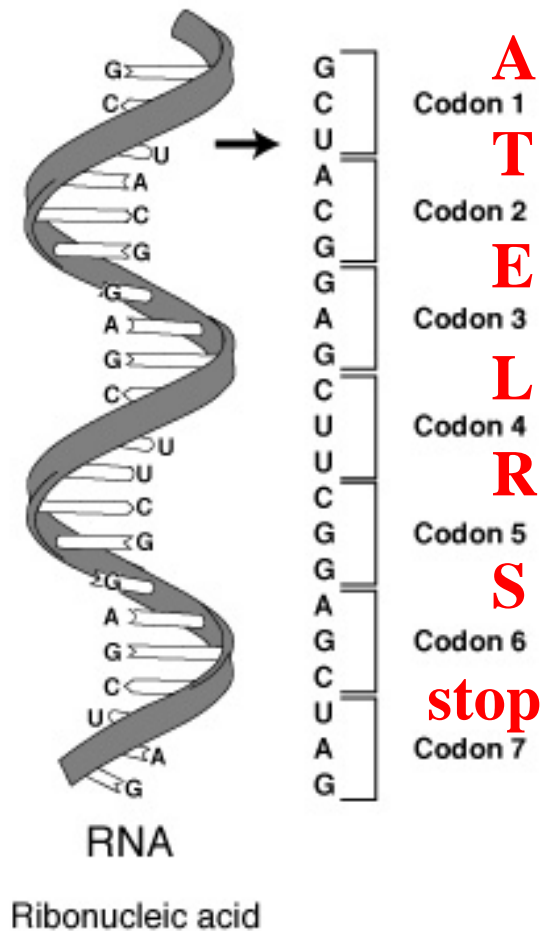


Splicing: nRNA → mRNA



RNA synthesis and processing

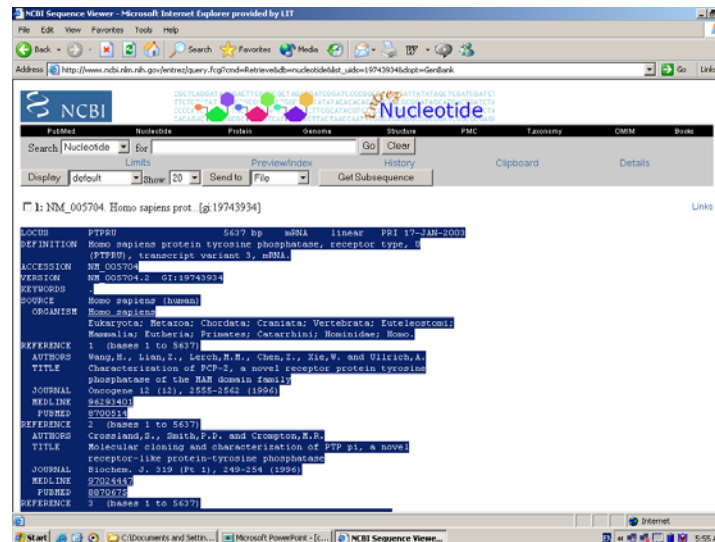
Translation: mRNA → protein



First	U	C	A	G	Last
U	Phe F	Ser S	Tyr Y	Cys C	U
	Phe	Ser	Tyr	Cys	C
	Leu L	Ser	Stop (Ochre)	Stop (Umber)	A
	Leu	Ser	Stop (Amber)	Trp W	G
C	Leu	Pro P	His H	Arg R	U
	Leu	Pro	His	Arg	C
	Leu	Pro	Gln Q	Arg	A
	Leu	Pro	Gln	Arg	G
A	Ile I	Thr T	Asn N	Ser	U
	Ile	Thr	Asn	Ser	C
	Ile	Thr	Lys K	Arg	A
	Met M	Thr	Lys	Arg	G
G	Val V	Ala A	Asp D	Gly G	U
	Val	Ala	Asp	Gly	C
	Val	Ala	Glu E	Gly	A
	Val	Ala	Glu	Gly	G

What does DNA data look like?

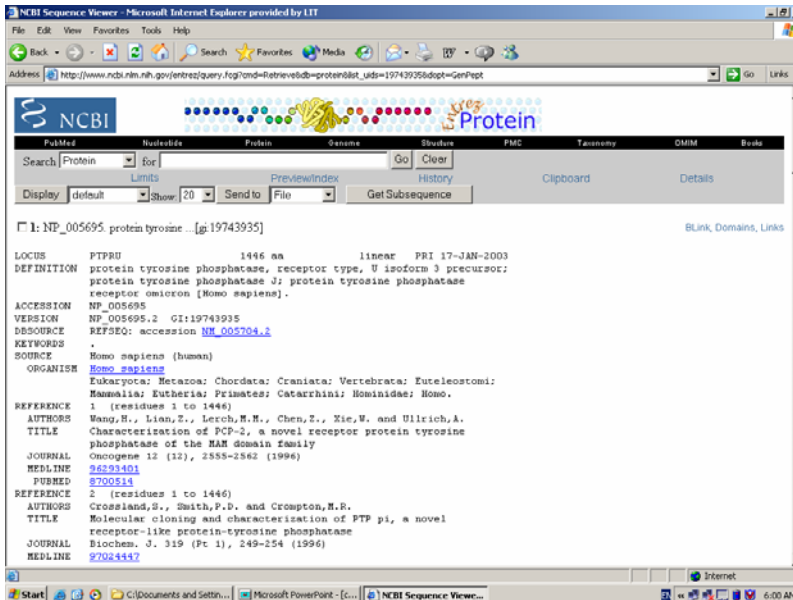
- A sample GenBank record from NCBI
- http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=nucleotide&list_uids=19743934&dopt=GenBank



```
NCBI Sequence Viewer - Microsoft Internet Explorer provided by LIT
Address: http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=nucleotide&list_uids=19743934&dopt=GenBank
Nucleotide
Search: Nucleotide
Display: default
1: NM_005704 Homo sapiens prot. [gi:19743934]
LOCUS       PTP2      5637 bp     mRNA     linear     P1 17-108-100
DEFINITION Homo sapiens protein tyrosine phosphatase, receptor type, 2
             (PTP2), transcript variant 3, mRNA.
ACCESSION   NM_005704
VERSION     NM_005704.2  GI:19743934
KEYWORDS   .
SOURCE      Homo sapiens (human)
ORGANISM    Homo sapiens
             Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi;
             Mammalia; Eutheria; Primates; Catarrhini; Hominidae; Homo.
REFERENCE   1  (bases 1 to 5637)
AUTHORS    Wang, H., Lim, S., Seck, R.H., Chen, Z., Xie, Y. and Ullrich, A.
TITLE      Characterization of PCP-2, a novel receptor protein tyrosine
             phosphatase of the SH-PTP family.
JOURNAL    Oncogene 13 (12), 2555-2562 (1996)
MEDLINE    8628140
PubMed    8700511
REFERENCE   2  (bases 1 to 5637)
AUTHORS    Crossland, S., Smith, P.D. and Crompton, R.B.
TITLE      Molecular cloning and characterization of PTP gi, a novel
             receptor-like protein-tyrosine phosphatase.
JOURNAL    Biochem. J. 316 (Pt 3), 849-854 (1996)
MEDLINE    8704447
PubMed    8876577
REFERENCE   ?  (bases 1 to 5637)
```


What does protein data look like?

- A sample GenPept record from NCBI
- http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=protein&list_uids=19743935&dopt=GenPept



NCBI Sequence Viewer - Microsoft Internet Explorer provided by LIT

Address: http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=protein&list_uids=19743935&dopt=GenPept

NCBI Protein

Search: Protein for [] Go Clear

Display: default Show: 20 Send to: File Get Subsequence

1: NP_005695. protein tyrosine ... [gi:19743935]

LOCUS FTPTJ 1446 aa linear PRI 17-JAN-2003
DEFINITION protein tyrosine phosphatase, receptor type, II isoform 3 precursor;
protein tyrosine phosphatase J; protein tyrosine phosphatase
receptor omicron [Homo sapiens].

ACCESSION NP_005695
VERSION NP_005695.2 GI:19743935
DBSOURCE REFSEQ: accession [NR_005704.2](#)

KEYWORDS .

SOURCE Homo sapiens (human)

ORGANISM [Homo_sapiens](#)
Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi;
Mammalia; Eutheria; Primates; Catarrhini; Hominidae; Homo.

REFERENCE
1 (residues 1 to 1446)
AUTHORS Wang, H., Lion, Z., Lerch, M.M., Chen, Z., Xie, V. and Ulrich, A.
TITLE Characterization of PCP-2, a novel receptor protein tyrosine
phosphatase of the SHM domain family
JOURNAL Oncogene 12 (12), 2555-2562 (1996)
MEDLINE [96391401](#)
PUBMED [8700514](#)

REFERENCE
2 (residues 1 to 1446)
AUTHORS Crossland, S., Smith, P.D. and Crompton, M.R.
TITLE Molecular cloning and characterization of PTP pi, a novel
receptor-like protein-tyrosine phosphatase
JOURNAL Biochem. J. 319 (Pt 1), 249-254 (1996)
MEDLINE [87064447](#)

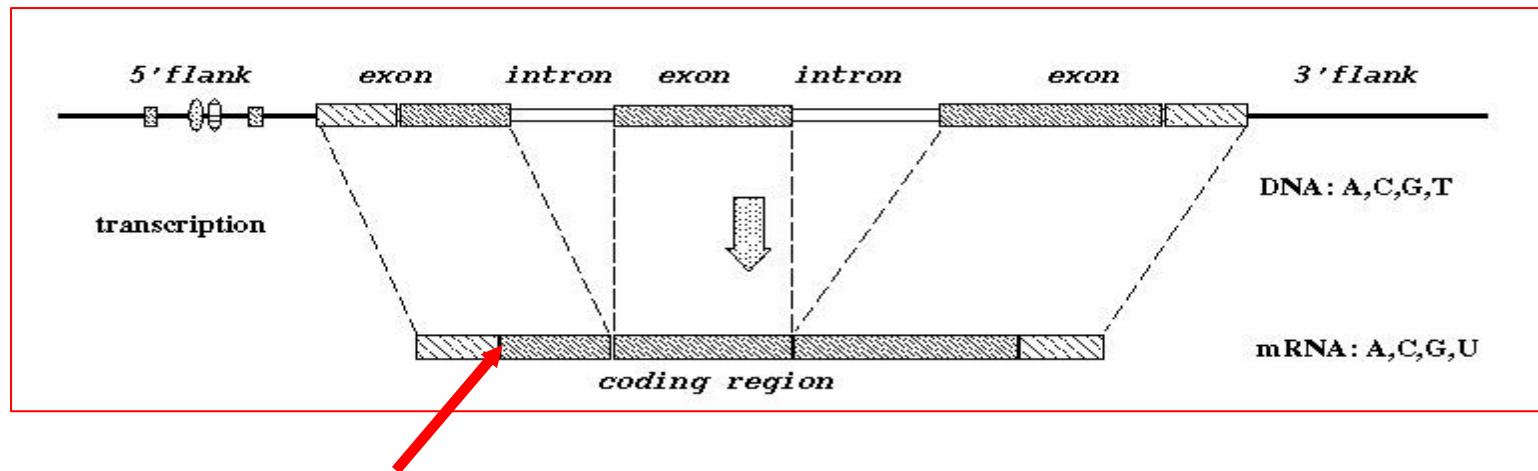


Recognition of Translation Initiation Sites

**An introduction to the World's simplest TIS
recognition system**



Translation Initiation Site



A Sample cDNA

```

299 HSU27655.1 CAT U27655 Homo sapiens
CGTGTGTGCAGCAGCCTGCAGCTGCCCAAGCCATGGCTGAACACTGACTCCCAGCTGTG      80
CCCAGGGCTTCAAAGACTTCTCAGCTTCGAGCATGGCTTTTGGCTGTCAGGGCAGCTGTA      160
GGAGGCAGATGAGAAGAGGGAGATGGCCTTGGAGGAAGGGAAGGGGCCTGGTGCCGAGGA      240
CCTCTCCTGGCCAGGAGCTTCCTCCAGGACAAGACCTTCCACCCAACAAGGACTCCCCT
.....                                                                    80
.....iEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEE      160
EEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEE      240
EEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEEE
  
```

- What makes the second ATG the TIS?

Approach

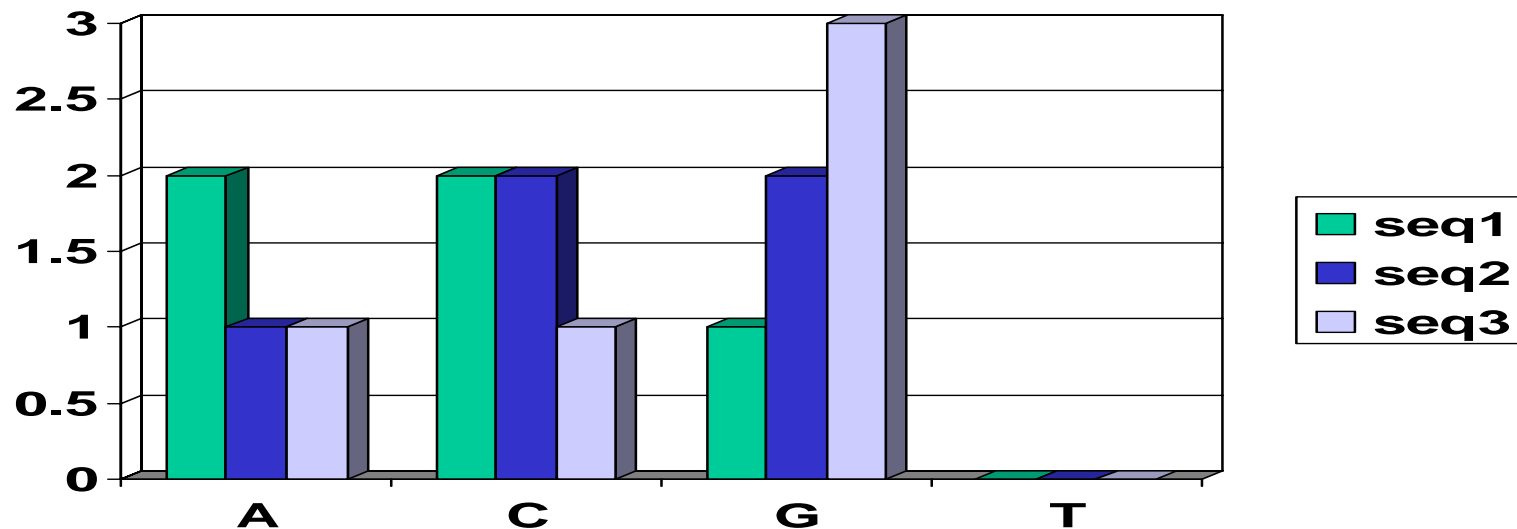
- **Training data gathering**
- **Signal generation**
 - k-grams, distance, domain know-how, ...
- **Signal selection**
 - Entropy, χ^2 , CFS, t-test, domain know-how...
- **Signal integration**
 - SVM, ANN, PCL, CART, C4.5, kNN, ...

Training & Testing Data

- **Vertebrate dataset of Pedersen & Nielsen [ISMB'97]**
- **3312 sequences**
- **13503 ATG sites**
- **3312 (24.5%) are TIS**
- **10191 (75.5%) are non-TIS**
- **Use for 3-fold x-validation expts**

Signal Generation

- **K-grams (ie., k consecutive letters)**
 - $K = 1, 2, 3, 4, 5, \dots$
 - Window size vs. fixed position
 - Up-stream, downstream vs. any where in window
 - In-frame vs. any frame



Signal Generation: An Example

```

299 HSU27655.1 CAT U27655 Homo sapiens
CGTGTGTGCAGCAGCCTGCAGCTGCCCAAGCCATGGCTGAACACTGACTCCCAGCTGTG      80
CCCAGGGCTTCAAAGACTTCTCAGCTTCGAGCATGGCTTTTGGCTGTCAGGGCAGCTGTA    160
GGAGGCAGATGAGAAGAGGGAGATGGCCTTGGAGGAAGGGAAGGGCCTGGTGCCGAGGA    240
CCTCTCCTGGCCAGGAGCTTCCTCCAGGACAAGACCTTCCACCCAACAAGGACTCCCCT
  
```

- **Window = ± 100 bases**
- **In-frame, downstream**
 - GCT = 1, TTT = 1, ATG = 1...
- **Any-frame, downstream**
 - GCT = 3, TTT = 2, ATG = 2...
- **In-frame, upstream**
 - GCT = 2, TTT = 0, ATG = 0, ...

Exercise: Find the in-frame downstream ATG

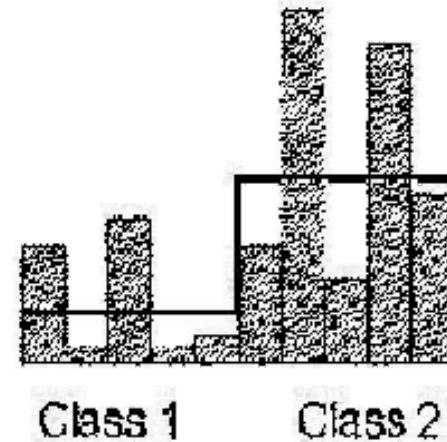
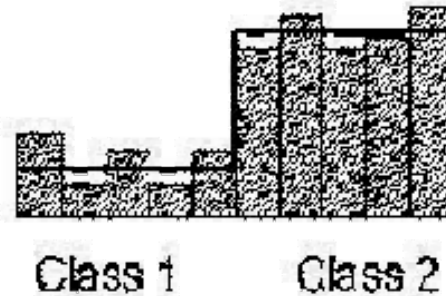
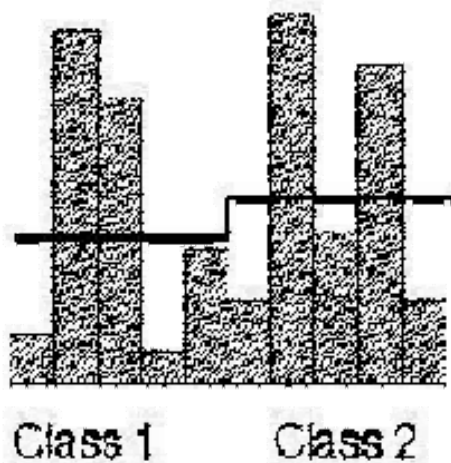
Exercise: What are the possible k-grams (k=3) in this sequence?

Too Many Signals

- For each value of k , there are $4^k * 3 * 2$ k -grams
- If we use $k = 1, 2, 3, 4, 5$, we have $24 + 96 + 384 + 1536 + 6144 = 8184$ features!
- This is too many for most machine learning algorithms

Signal Selection (Basic Idea)

- Choose a signal w/ low intra-class distance
- Choose a signal w/ high inter-class distance



Signal Selection (e.g., t-statistics)

The t-stats of a signal is defined as

$$t = \frac{|\mu_1 - \mu_2|}{\sqrt{(\sigma_1^2/n_1) + (\sigma_2^2/n_2)}}$$

where σ_i^2 is the variance of that signal in class i , μ_i is the mean of that signal in class i , and n_i is the size of class i .

Signal Selection (e.g., MIT-correlation)

The MIT-correlation value of a signal is defined as

$$MIT = \frac{|\mu_1 - \mu_2|}{\sigma_1 + \sigma_2}$$

where σ_i is the standard deviation of that signal in class i and μ_i is the mean of that signal in class i .

Signal Selection (e.g., χ^2)

The χ^2 value of a signal is defined as:

$$\chi^2 = \sum_{i=1}^m \sum_{j=1}^k \frac{(A_{ij} - E_{ij})^2}{E_{ij}},$$

where m is the number of intervals, k the number of classes, A_{ij} the number of samples in the i th interval, j th class, R_i the number of samples in the i th interval, C_j the number of samples in the j th class, N the total number of samples, and E_{ij} the expected frequency of A_{ij} ($E_{ij} = R_i * C_j / N$).

Signal Selection (e.g., CFS)

- **Instead of scoring individual signals, how about scoring a group of signals as a whole?**
- **CFS**
 - Correlation-based Feature Selection
 - A good group contains signals that are highly correlated with the class, and yet uncorrelated with each other

Exercise: What is the main challenge in implementing CFS?

Sample k-grams Selected by CFS for Recognizing TIS



Kozak consensus

Leaky scanning

Stop codon

- **Position -3**
- **in-frame upstream ATG**
- **in-frame downstream**
 - TAA, TAG, TGA,
 - CTG, GAC, GAG, and GCC

Codon bias?

Signal Integration

- **kNN**
 - Given a test sample, find the k training samples that are most similar to it. Let the majority class win
- **SVM**
 - Given a group of training samples from two classes, determine a separating plane that maximises the margin of error
- **Naïve Bayes, ANN, C4.5, ...**

Results (3-fold x-validation)

	predicted as positive	predicted as negative
positive	TP	FN
negative	FP	TN

Exercise:
What is $TP/(TP+FP)$?

	$TP/(TP + FN)$	$TN/(TN + FP)$	$TP/(TP + FP)$	Accuracy
Naïve Bayes	84.3%	86.1%	66.3%	85.7%
SVM	73.9%	93.2%	77.9%	88.5%
Neural Network	77.6%	93.2%	78.8%	89.4%
Decision Tree	74.0%	94.4%	81.1%	89.4%

Improvement by Voting

- Apply any 3 of Naïve Bayes, SVM, Neural Network, & Decision Tree. Decide by majority

	TP/(TP + FN)	TN/(TN + FP)	TP/(TP + FP)	Accuracy
NB+SVM+NN	79.2%	92.1%	76.5%	88.9%
NB+SVM+Tree	78.8%	92.0%	76.2%	88.8%
NB+NN+Tree	77.6%	94.5%	82.1%	90.4%
SVM+NN+Tree	75.9%	94.3%	81.2%	89.8%
Best of 4	84.3%	94.4%	81.1%	89.4%
Worst of 4	73.9%	86.1%	66.3%	85.7%

Improvement by Scanning

- Apply Naïve Bayes or SVM left-to-right until first ATG predicted as positive. That's the TIS
- Naïve Bayes & SVM models were trained using TIS vs. Up-stream ATG

	TP/(TP + FN)	TN/(TN + FP)	TP/(TP + FP)	Accuracy
NB	84.3%	86.1%	66.3%	85.7%
SVM	73.9%	93.2%	77.9%	88.5%
NB+Scanning	87.3%	96.1%	87.9%	93.9%
SVM+Scanning	88.5%	96.3%	88.6%	94.4%

Performance Comparisons

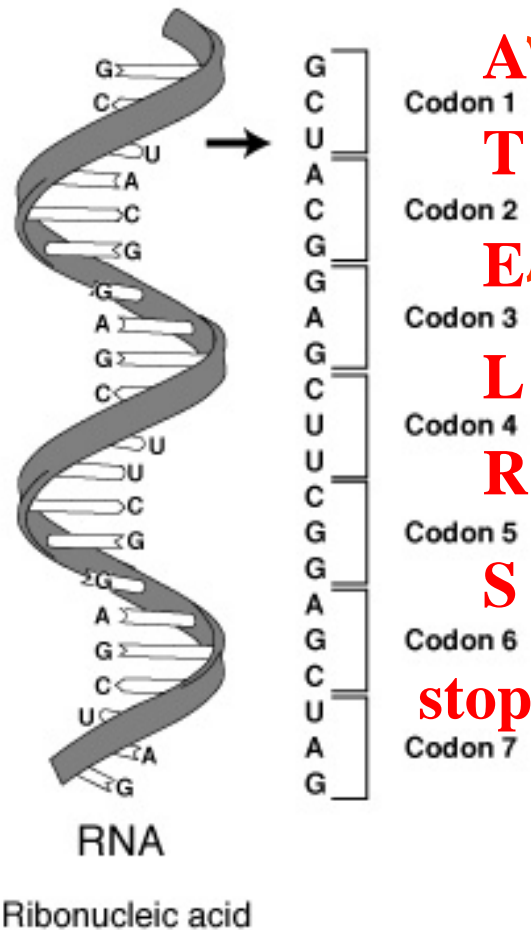
	TP/(TP + FN)	TN/(TN + FP)	TP/(TP + FP)	Accuracy
NB	84.3%	86.1%	66.3%	85.7%
Decision Tree	74.0%	94.4%	81.1%	89.4%
NB+NN+Tree	77.6%	94.5%	82.1%	90.4%
SVM+Scanning	88.5%	96.3%	88.6%	94.4%*
Pedersen&Nielsen	78%	87%	-	85%
Zien	69.9%	94.1%	-	88.1%
Hatzigeorgiou	-	-	-	94%*

* result not directly comparable

Technique Comparisons

- **Pedersen&Nielsen [ISMB'97]**
 - Neural network
 - No explicit features
- **Zien [Bioinformatics'00]**
 - SVM+kernel engineering
 - No explicit features
- **Hatzigeorgiou [Bioinformatics'02]**
 - Multiple neural networks
 - Scanning rule
 - No explicit features
- **Our approach**
 - Explicit feature generation
 - Explicit feature selection
 - Use any machine learning method w/o any form of complicated tuning
 - Scanning rule is optional

mRNA → protein

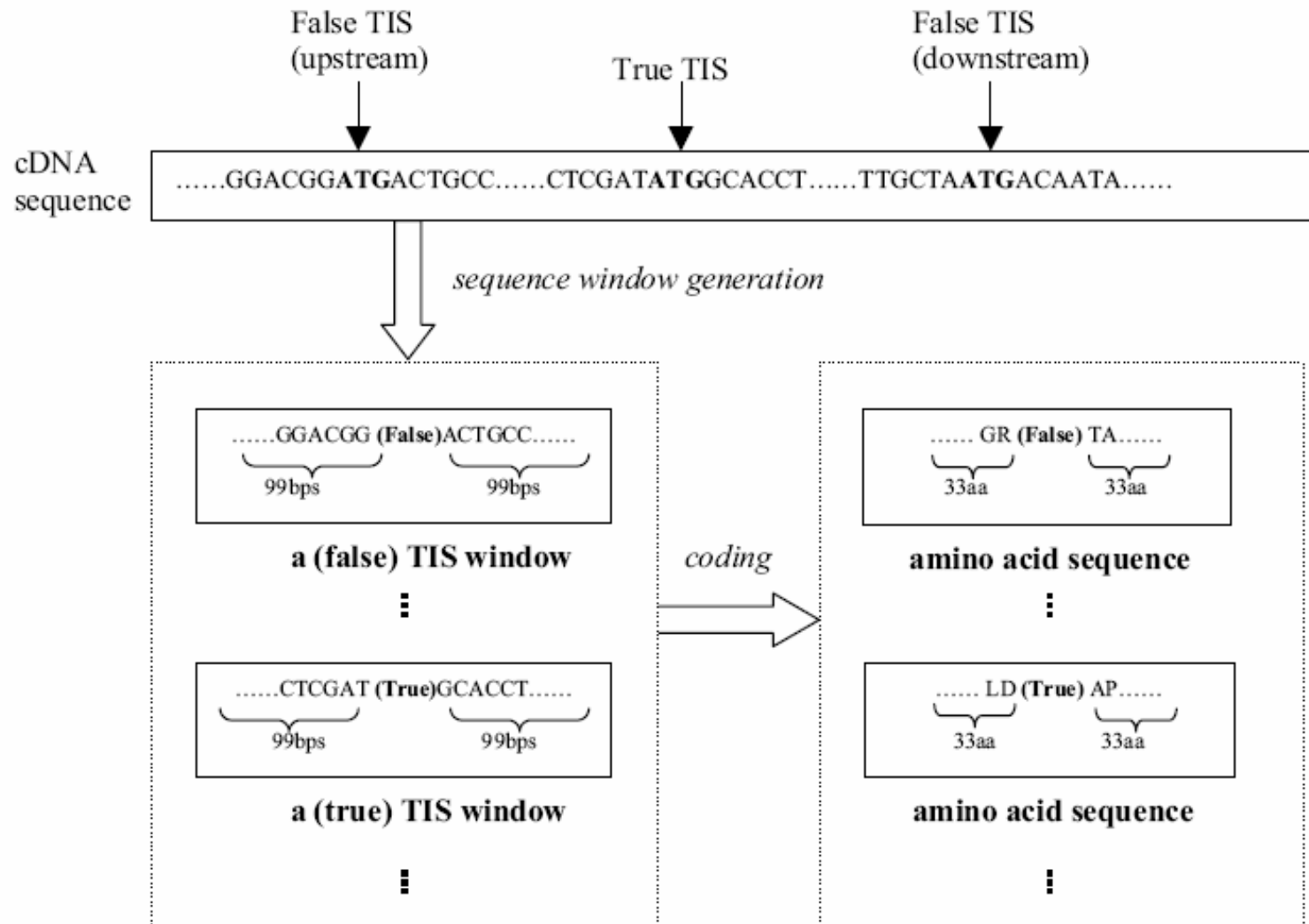


How about using k-grams from the translation?

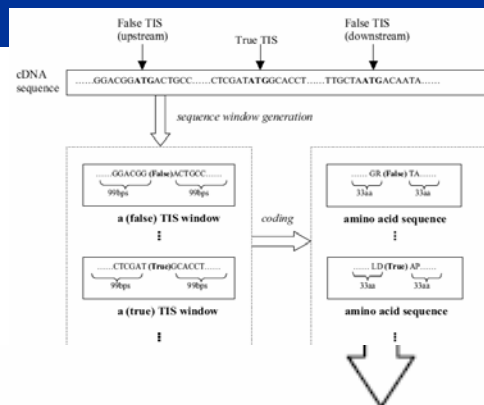
First	U	C	A	G	Last
U	Phe F	Ser S	Tyr Y	Cys C	U
	Phe F	Ser S	Tyr Y	Cys C	C
	Leu L	Ser	Stop (Ochre)	Stop (Umber)	A
	Leu L	Ser	Stop (Amber)	Trp W	G
C	Leu	Pro P	His H	Arg R	U
	Leu	Pro P	His H	Arg R	C
	Leu	Pro	Gln Q	Arg R	A
	Leu	Pro	Gln Q	Arg R	G
A	Ile I	Thr T	Asn N	Ser	U
	Ile I	Thr T	Asn N	Ser	C
	Ile	Thr	Lys K	Arg	A
	Met M	Thr	Lys K	Arg	G
G	Val V	Ala A	Asp D	Gly G	U
	Val V	Ala A	Asp D	Gly G	C
	Val	Ala	Glu E	Gly G	A
	Val	Ala	Glu E	Gly G	G

Exercise: List the first 10 amino acid in our example sequence

Amino-Acid Features

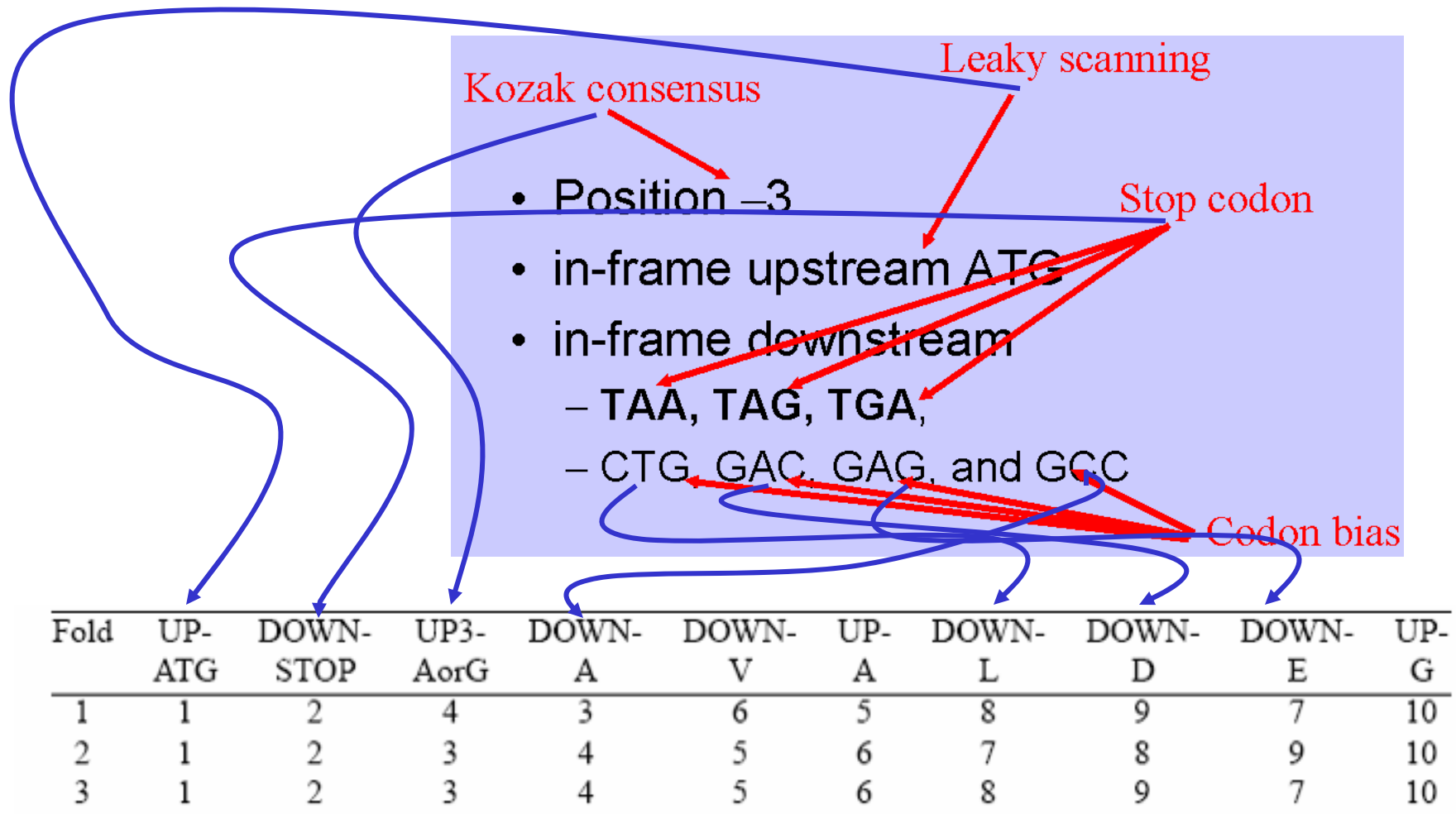


Amino-Acid Features



New feature space (total of 927 features + class label)			
42 1-gram amino acid patterns	882 2-gram amino acid patterns	3 bio-know-ledge patterns	class label
UP-A, UP-R, ...,UP-N, DOWN-A, DOWN-R, ..., DOWN-N (numeric type)	UP-AA, UP-AR, ..., UP-NN, DOWN-AA, DOWN-AR, ..., DOWN-NN (numeric type)	DOWN4-G UP3-AorG, UP-ATG (boolean type, Y or N)	True, False
Frequency as values			
1, 3, 5, 0, 4, ... ⋮	6, 2, 7, 0, 5, ... ⋮	N, N, N, ⋮	False ⋮
6, 5, 7, 9, 0, ... ⋮	2, 0, 3, 10, 0, ... ⋮	Y, Y, Y, ⋮	True ⋮

Amino Acid K-grams Discovered (by entropy)



Independent Validation Sets

- **A. Hatzigeorgiou:**
 - 480 fully sequenced human cDNAs
 - 188 left after eliminating sequences similar to training set (Pedersen & Nielsen's)
 - 3.42% of ATGs are TIS
- **Our own:**
 - well characterized human gene sequences from chromosome X (565 TIS) and chromosome 21 (180 TIS)

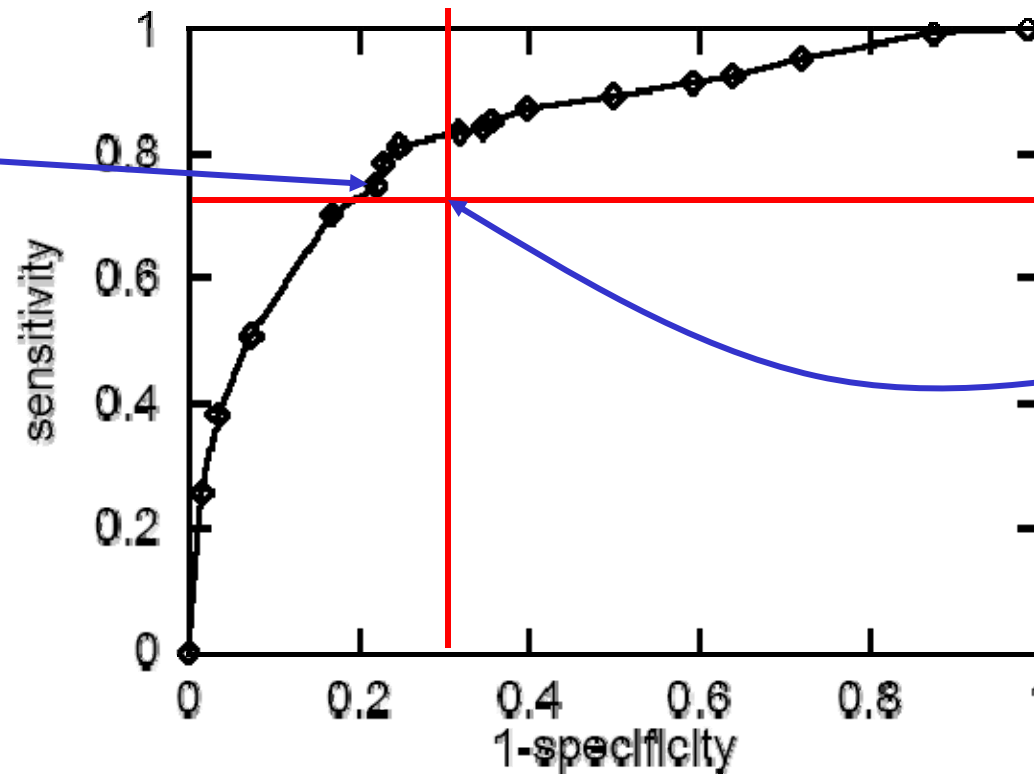
Validation Results (on Hatzigeorgiou's)

Algorithm	Sensitivity	Specificity	Precision	Accuracy
SVMs(linear)	96.28%	89.15%	25.31%	89.42%
SVMs(quad)	94.14%	90.13%	26.70%	90.28%
Ensemble Trees	92.02%	92.71%	32.52%	92.68%

- Using top 100 features selected by entropy and trained on Pedersen & Nielsen's dataset

Validation Results (on Chr X and Chr 21)

Our
method



ATGpr

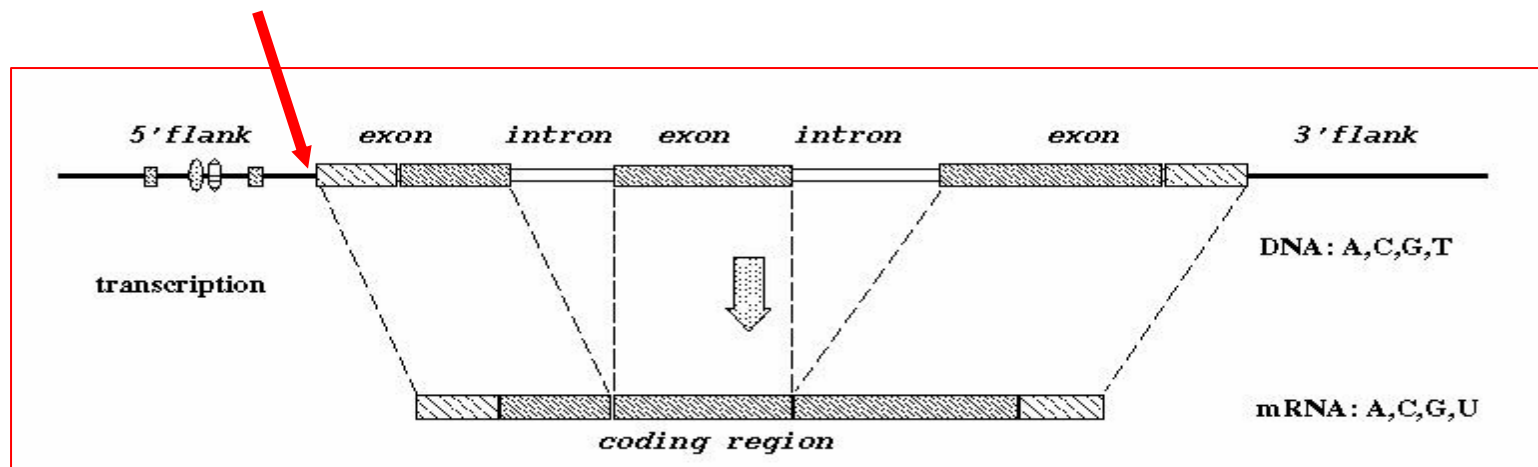
- Using top 100 features selected by entropy and trained on Pedersen & Nielsen's

Recognition of Transcription Start Sites

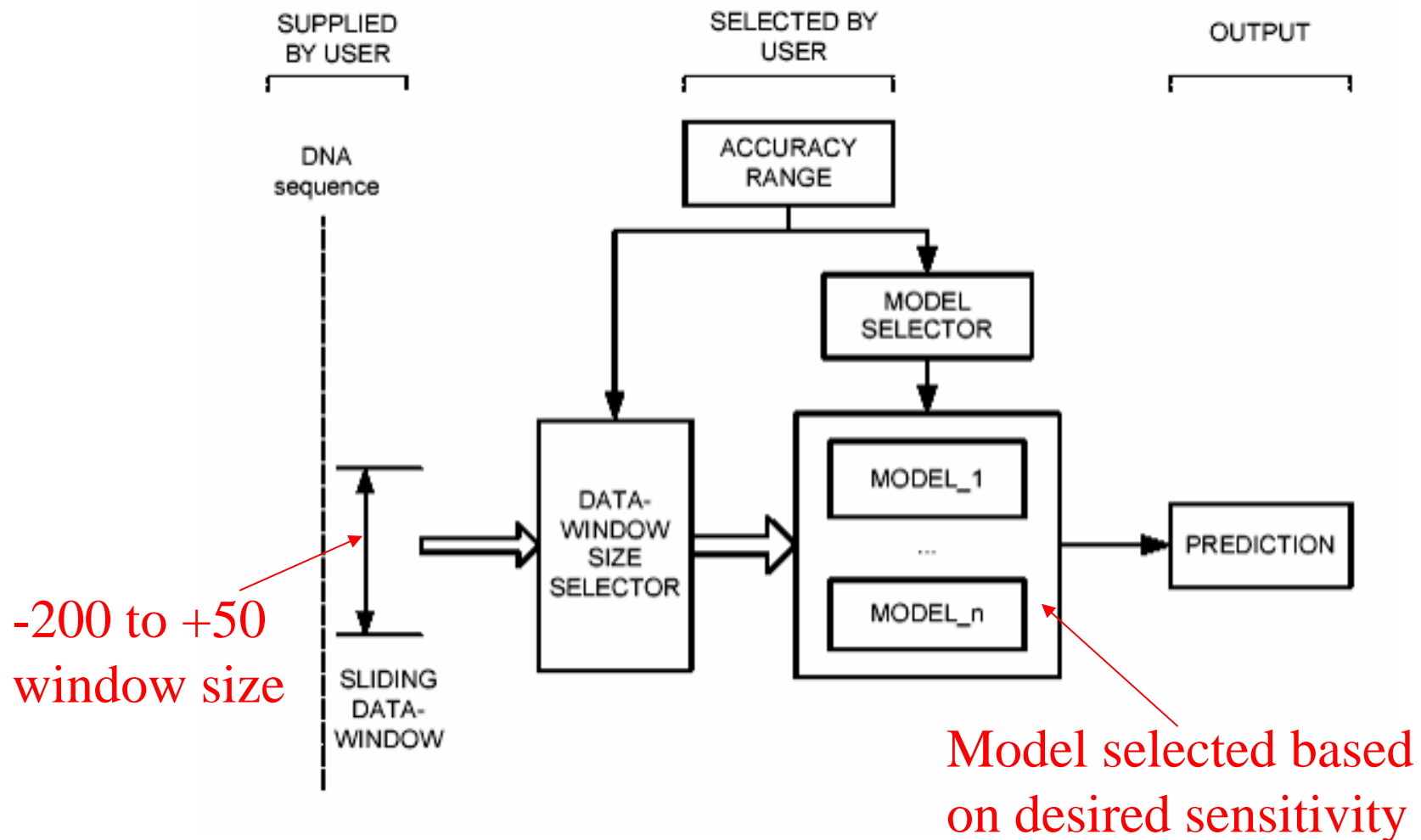
**An introduction to the World's best TSS
recognition system:
A heavy tuning approach**



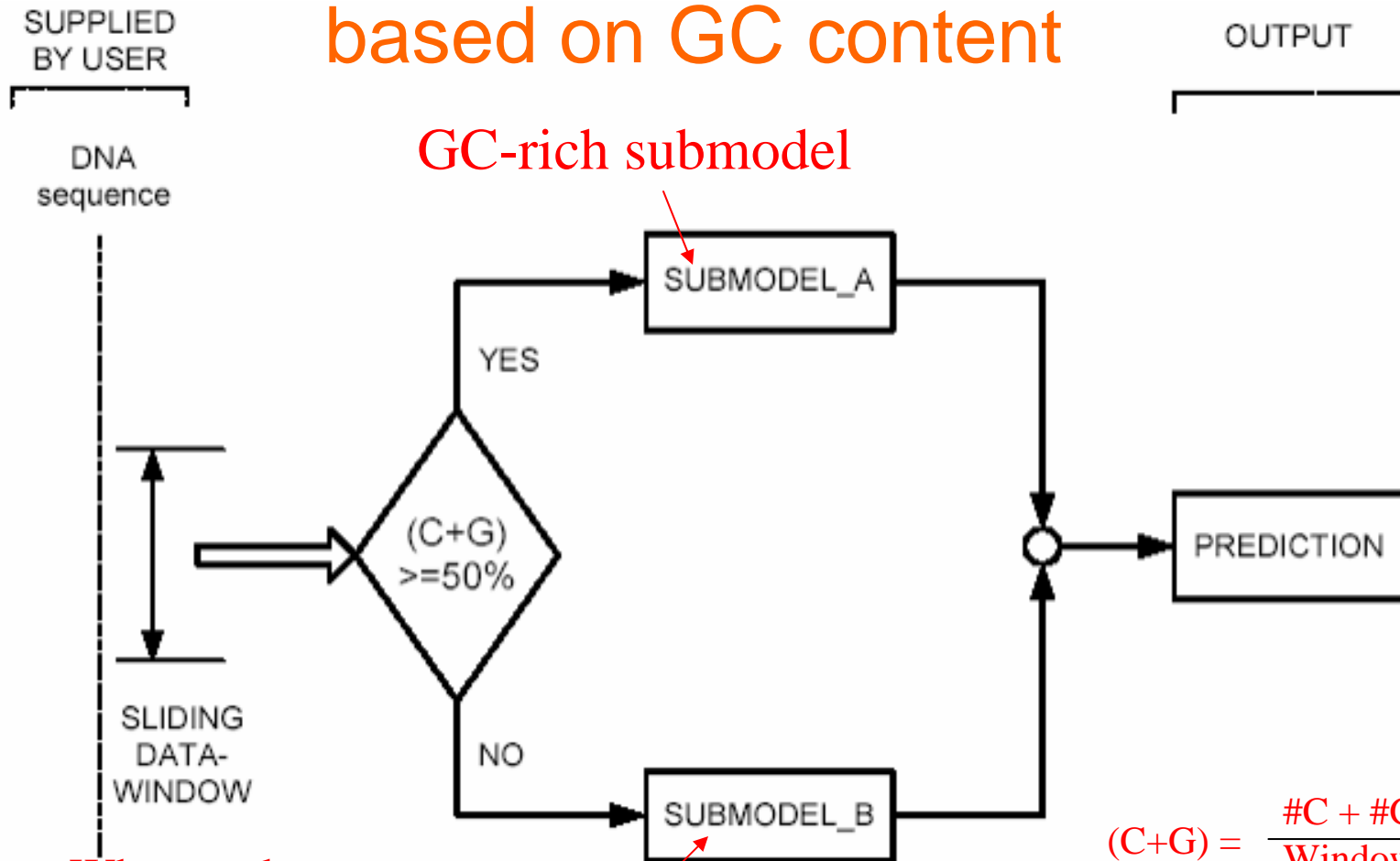
Transcription Start Site



Structure of Dragon Promoter Finder

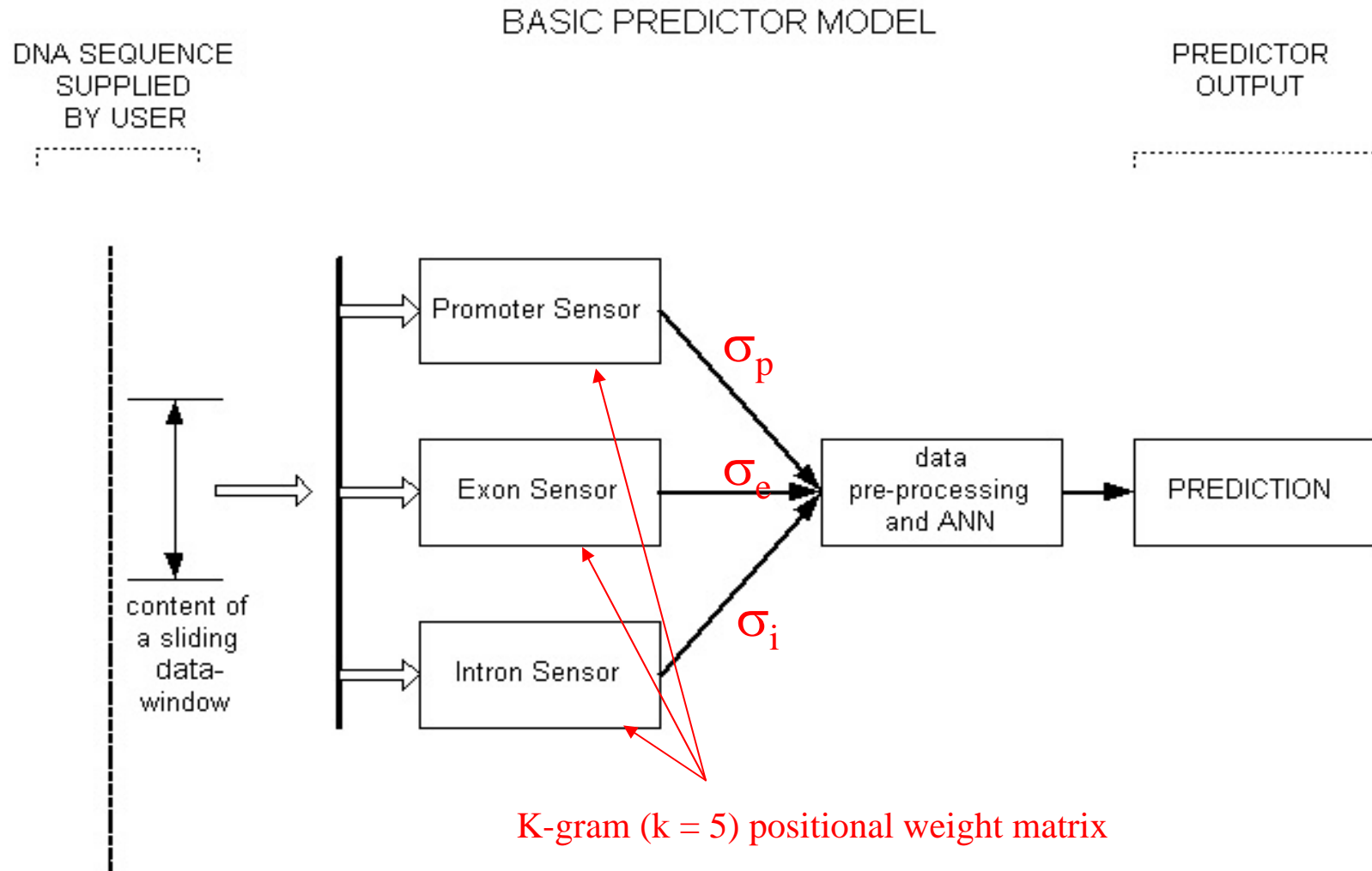


Each model has two submodels based on GC content



Exercise: Why are the submodels based on GC content?

Data Analysis Within Submodel



Promoter, Exon, Intron Sensors

- These sensors are positional weight matrices of k-grams, k = 5 (aka pentamers)
- They are calculated as below using promoter, exon, intron data respectively

$$\sigma = \frac{\left(\sum_{i=1}^{L-4} p_j^i \otimes f_{j,i} \right)}{\left(\sum_{i=1}^{L-4} \max_j f_{j,i} \right)}, \quad p_j^i \otimes f_{j,i} = \begin{cases} f_{j,i}, & \text{if } p_i = p_j^i \\ 0, & \text{if } p_i \neq p_j^i \end{cases}$$

Window size

Pentamer at i^{th} position in input

Frequency of j^{th} pentamer at i^{th} position in training window

j^{th} pentamer at i^{th} position in training window

Just to make sure you know what I mean ...

- Give me 3 DNA seq of length 10:
 - Seq₁ = ACCGAGTTCT
 - Seq₂ = AGTGTACCTG
 - Seq₃ = AGTTCGTATG
- Then

1-mer	pos1	pos2	pos3	pos4	pos5	pos6	pos7	pos8	pos9	pos10
A	3/3	0/3	0/3							
C	0/3	1/3	1/3		Exercise: Fill in the rest of the table					
G	0/3	2/3	0/3							
T	0/3	0/3	2/3							

Just to make sure you know what I mean ...

- Give me 3 DNA seq of length 10:
 - Seq₁ = ACCGAGTTCT
 - Seq₂ = AGTGTACCTG
 - Seq₃ = AGTTCGTATG
- Then

Exercise: How many rows should this 2-mer table have? How many rows should the pentamer table have?

2-mer	pos1	pos2	pos3	pos4	pos5	pos6	pos7	pos8	pos9
AA	0/3	0/3	0/3						
AC	1/3	0/3	0/3		Exercise: Fill in the rest of the table				
...						
TT	0/3	0/3	1/3				1/3		

Data Preprocessing & ANN

Tuning parameters

$$s_E = \text{sat}(\sigma_p - \sigma_e, a_e, b_e)$$

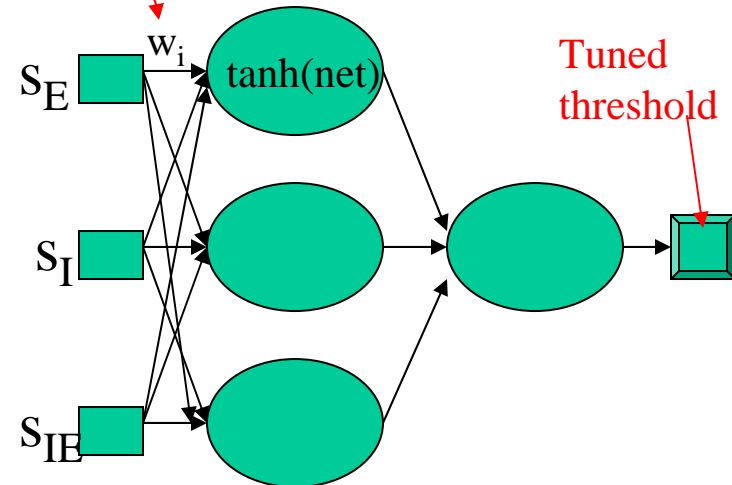
$$s_I = \text{sat}(\sigma_p - \sigma_i, a_i, b_i)$$

$$s_{EI} = \text{sat}(\sigma_e - \sigma_i, a_{ei}, b_{ei})$$

where the function *sat* is defined by

$$\text{sat}(x, a, b) = \begin{cases} a, & \text{if } x > a \\ x, & \text{if } b \leq x \leq a. \\ b, & \text{if } b > x \end{cases}$$

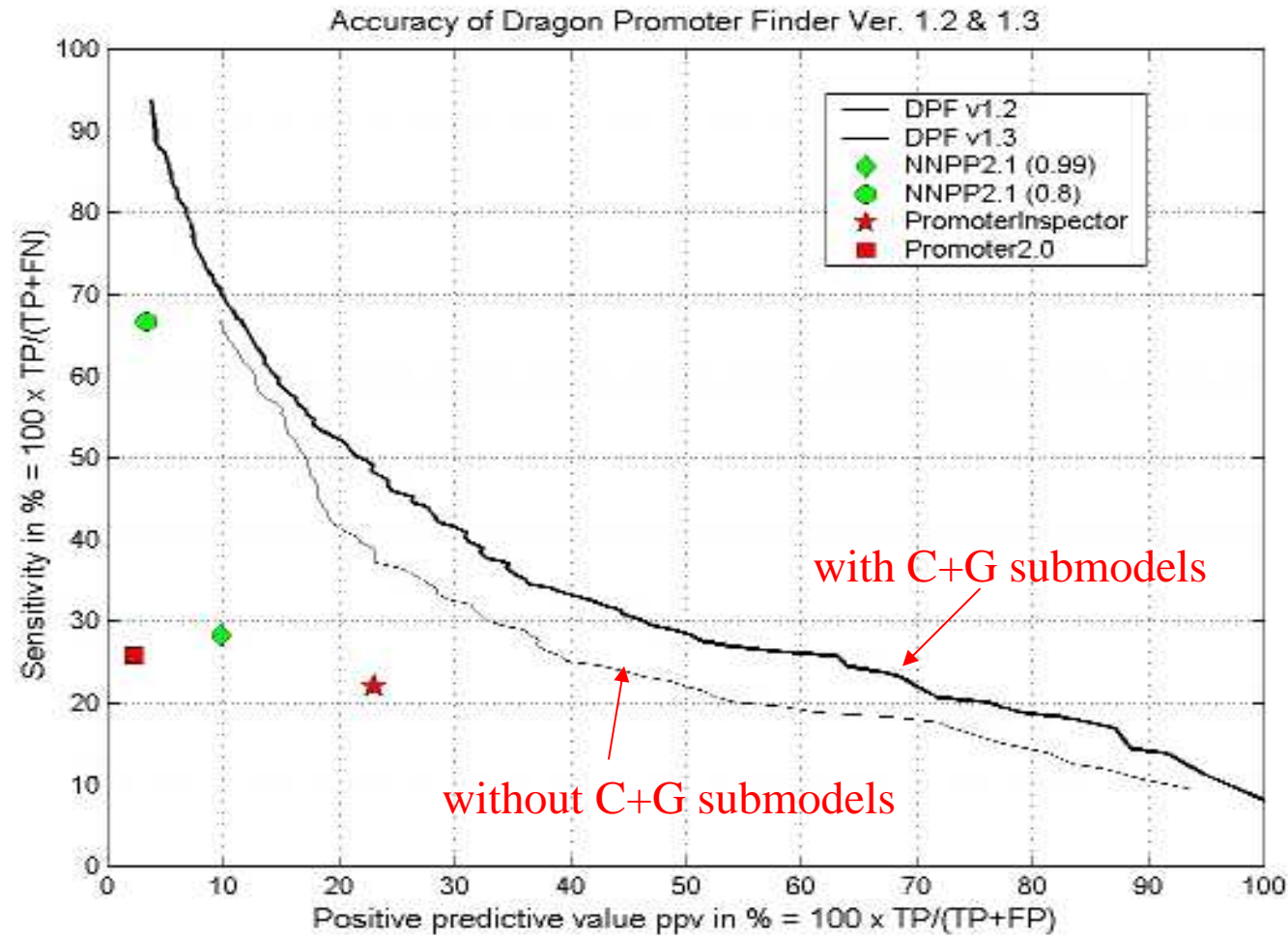
Simple feedforward ANN
 trained by the Bayesian
 regularisation method



$$\tanh(x) = \frac{e^x - e^{-x}}{e^x + e^{-x}}$$

$$\text{net} = \sum s_i * w_i$$

Accuracy Comparisons



Training Data Criteria & Preparation

- **Contain both positive and negative sequences**
- **Sufficient diversity, resembling different transcription start mechanisms**
- **Sufficient diversity, resembling different non-promoters**
- **Sanitized as much as possible**
- **TSS taken from**
 - 793 vertebrate promoters from EPD
 - -200 to +50 bp of TSS
- **non-TSS taken from**
 - GenBank,
 - 800 exons
 - 4000 introns,
 - 250 bp,
 - non-overlapping,
 - <50% identities

Tuning Data Preparation

- **To tune adjustable system parameters in Dragon, we need a separate tuning data set**
- **TSS taken from**
 - 20 full-length gene seqs with known TSS
 - -200 to +50 bp of TSS
 - no overlap with EPD
- **Non-TSS taken from**
 - 1600 human 3'UTR seqs
 - 500 human exons
 - 500 human introns
 - 250 bp
 - no overlap

Testing Data Criteria & Preparation

- Seqs should be from the training or evaluation of other systems (no bias!)
- Seqs should be disjoint from training and tuning data sets
- Seqs should have TSS
- Seqs should be cleaned to remove redundancy, <50% identities
- 159 TSS from 147 human and human virus seqs
- cumulative length of more than 1.15Mbp
- Taken from GENESCAN, Geneld, Genie, etc.

Accuracy on Human Chromosome

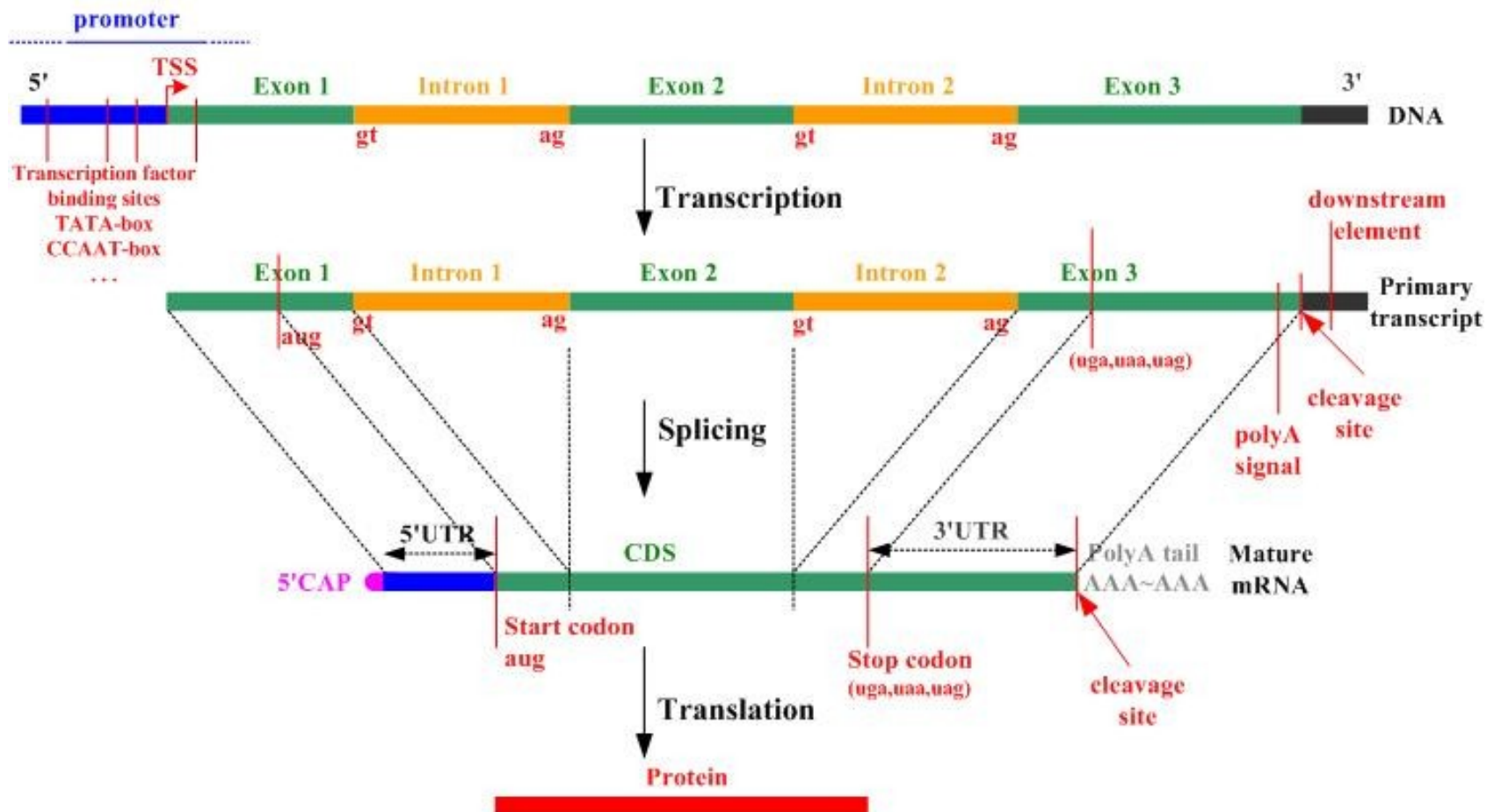


Human chromosome 22 (known genes)	
Se	Ppv
49%	48%
58%	42%
64%	33%
74%	30%
80%	23%

Other Gene Features



Other Gene Features



Any Question?



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